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(54) Title: GROWTH DIFFERENTIATION FACTOR-9						
(57) Abstract						
Growth differentiation factor-9 (GDF-9) is disclosed along with its polynucleotide sequence and amino acid sequence. Also disclosed are diagnostic and therapeutic methods of using the GDF-9 polypeptide and polynucleotide sequences.						
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GROWTH DIFFERENTIATION FACTOR-9

This application is a continuation-in-part application of U.S. Serial No. 08/003,303, filed January 12, 1993.

BACKGROUND OF THE INVENTION

1. Field of the Invention

The invention relates generally to growth factors and specifically to a new member of the transforming growth factor beta (TGF- β) superfamily, which is denoted, growth differentiation factor-9 (GDF-9).

2. Description of Related Art

The transforming growth factor *β* (TGF-*β*) superfamily encompasses a group of structurally-related proteins which affect a wide range of differentiation processes during embryonic development. The family includes, Mullerian inhibiting substance (MIS), which is required for normal male sex development (Behringer, *et al.*, *Nature*, 345:167, 1990), *Drosophila* decapentaplegic (DPP) gene product, which is required for dorsal-ventral axis formation and morphogenesis of the imaginal disks (Padgett, *et al.*, *Nature*, 325:81-84, 1987), the *Xenopus* Vg-1 gene product, which localizes to the vegetal pole of eggs ((Weeks, *et al.*, *Cell*, 51:861-867, 1987), the activins (Mason, *et al.*, *Biochem, Biophys. Res. Commun.*, 135:957-964, 1986), which can induce the formation of mesoderm and anterior structures in *Xenopus* embryos (Thomsen, *et al.*, *Cell*, 63:485, 1990), and the bone morphogenetic proteins (BMPs, osteogenin, OP-1) which can induce *de novo* cartilage and bone formation (Sampath, *et al.*, *J. Biol. Chem.*, 265:13198, 1990). The TGF-*β*s can influence a variety of differentiation processes, including adipogenesis, myogenesis, chondrogenesis,

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hematopoiesis, and epithelial cell differentiation (for review, see Massague, *Cell* 49:437, 1987).

The proteins of the TGF- β family are initially synthesized as a large precursor protein which subsequently undergoes proteolytic cleavage at a cluster of basic residues approximately 110-140 amino acids from the C-terminus. The C-terminal regions of the proteins are all structurally related and the different family members can be classified into distinct subgroups based on the extent of their homology. Although the homologies within particular subgroups range from 70% to 90% amino acid sequence identity, the homologies between subgroups are significantly lower, generally ranging from only 20% to 50%. In each case, the active species appears to be a disulfide-linked dimer of C-terminal fragments. For most of the family members that have been studied, the homodimeric species has been found to be biologically active, but for other family members, like the inhibins (Ling, et al., Nature, 321:779, 1986) and the TGF- β s (Cheifetz, et al., Cell, 48:409, 1987), heterodimers have also been detected, and these appear to have different biological properties than the respective homodimers.

The inhibins and activins were originally purified from follicular fluid and shown to have counteracting effects on the release of follicle-stimulating hormone by the pituitary gland. Although the mRNAs for all three inhibin/activin subunits (αa, βA and βB) have been detected in the ovary, none of these appear to be ovary-specific (Meunier, et al., Proc.Natl.Acad.Sci. USA, 85:247, 1988). MIS has also been shown to be expressed by granulosa cells and the effects of MIS on ovarian development have been documented both *in vivo* in transgenic mice expressing MIS ectopically (Behringer, *supra*) and *in vitro* in organ culture (Vigier, et al., Development, 100:43, 1987).

Identification of new_factors_that are tissue-specific in their expression pattern will provide a greater understanding of that tissue's development and function.

SUMMARY OF THE INVENTION

The present invention provides a cell growth and differentiation factor, GDF-9, a polynucleotide sequence which encodes the factor and antibodies which are immunoreactive with the factor. This factor appears to relate to various cell proliferative disorders, especially those involving ovarian tumors, such as granulosa cell tumors.

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Thus, in one embodiment, the invention provides a method for detecting a cell proliferative disorder of ovarian origin and which is associated with GDF-9. In another embodiment, the invention provides a method of treating a cell proliferative disorder associated with abnormal levels of expression of GDF-9, by suppressing or enhancing GDF-9 activity.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1 shows expression of GDF-9 mRNA in adult tissues.

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FIGURE 2 shows nucleotide and predicted amino acid sequence of murine GDF-9. Consensus N-glycosylation signals are denoted by plain boxes. The putative tetrabasic processing sites are denoted by stippled boxes. The inframe termination codons upstream of the putative initiating ATG and the consensus polyadenylation signals are underlined. The poly A tails are not shown. Numbers indicate nucleotide position relative to the 5' end.

FIGURE 3 shows the alignment of the C-terminal sequences of GDF-9 with other members of the TGF- β family. The conserved cysteine residues are shaded. Dashes denote gaps introduced in order to maximize alignment.

FIGURE 4 shows amino acid homologies among the different members of the TGF- β superfamily. Numbers represent percent amino acid identities between each pair calculated from the first conserved cysteine to the C-terminus. Boxes represent homologies among highly-related members within particular subgroups.

FIGURE 5 shows the immunohistochemical localization of GDF-9 protein. Adjacent sections of an adult ovary were either stained with hematoxylin and eosin (FIGURE 5a) or incubated with immune (FIGURE 5b) or pre-immune (FIGURE 5c) serum at a dilution of 1:500. Anti-GDF-9 antiserum was prepared by expressing the C-terminal portion of murine GDF-9 (residues 308-441) in bacteria, excising GDF-9 protein from preparative SDS gels, and immunizing rabbits. Sites of antibody binding were visualized using the Vectastain ABC kit (Vector Labs).

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FIGURE 6 shows a comparison of the predicted amino acid sequences of murine (top lines) and human (bottom lines) GDF-9. Numbers represent amino acid positions relative to the N-termini. Vertical lines represent sequence identities. Dots represent gaps introduced in order to maximize the alignment. The clear box shows the predicted proteolytic processing sites. The shaded boxes show the cysteine residues in the mature region of the proteins. The bars at the bottom show a schematic of the pre-(clear) and mature (shaded) regions of GDF-9 with the percent sequence identities between the murine and human sequences shown below.

FIGURE 7 shows *in situ* hybridization to adult ovary sections using a GDF-9 RNA probe. [35S]-labeled anti-sense (FIGURE 7a and 7c) or sense (FIGURE 7 b and 7d) GDF-9 RNA probes were hybridized to adjacent paraffinembedded sections of ovaries fixed in 4% paraformaldehyde. Sections were dipped in photographic emulsion, exposed, developed, and then stained with hematoxylin and eosin. Two representative fields are shown.

FIGURE 8 shows *in situ* hybridization to a postnatal day 4 ovary section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 8a) or dark-field (FIGURE 8b) illumination.

FIGURE 9 shows *in situ* hybridization to postnatal day 8 ovary sections using an antisense (FIGURE 9a) or sense (FIGURE 9b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

FIGURE 10 shows *in situ* hybridization to adult oviduct sections using an antisense (FIGURE 10a) or sense (FIGURE 10b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

FIGURE 11 shows *in situ* hybridization to an adult oviduct (0.5 days following fertilization) section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 11a) or dark-field (FIGURE 11b) illumination.

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DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a growth and differentiation factor, GDF-9 and a polynucleotide sequence encoding GDF-9. Unlike other members of the $TGF-\beta$ superfamily, GDF-9 expression is highly tissue specific, being expressed in cells primarily in ovarian tissue. In one embodiment, the invention provides a method for detection of a cell proliferative disorder of the ovary, which is associated with GDF-9 expression. In another embodiment, the invention provides a method for treating a cell proliferative disorder associated with abnormal expression of GDF-9 by using an agent which suppresses or enhances GDF-9 activity.

The TGF- β superfamily consists of multifunctionally polypeptides that control proliferation, differentiation, and other functions in many cell types. Many of the peptides have regulatory, both positive and negative, effects on other peptide growth factors. The structural homology between the GDF-9 protein of this invention and the members of the TGF- β family, indicates that GDF-9 is a new member of the family of growth and differentiation factors. Based on the known activities of many of the other members, it can be expected that GDF-9 will also possess biological activities that will make it useful as a diagnostic and therapeutic reagent.

For example, another regulatory protein that has been found to have structural homology with TGF- β is inhibin, a specific and potent polypeptide inhibitor of the pituitary secretion of FSH. Inhibin has been isolated from ovarian follicular fluid. Because of its suppression of FSH, inhibin has potential to be used as a contraceptive in both males and females. GDF-9 may possess similar biological activity since it is also an ovarian specific peptide.Inhibin has also been shown to be useful as a marker for certain ovarian tumors (Lappohn, et al., N. Engl. J. Med., 321:790, 1989). GDF-9 may also be useful as a marker

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for identifying primary and metastatic neoplasms of ovarian origin. Similarly, GDF-9 may be useful as an indicator of developmental anomalies in prenatal screening procedures.

Another peptide of the TGF-β family is MIS, produced by the testis and responsible for the regression of the Mullerian ducts in the male embryo. MIS has been show to inhibit the growth of human ovarian cancer in nude mice (Donahoe, et al., Ann. Surg., 194:472, 1981). GDF-9 may function similarly and may, therefore, be useful as an anti-cancer agent, such as for the treatment of ovarian cancer.

GDF-9 may also function as a growth stimulatory factor and, therefore, be useful for the survival of various cell populations *in vitro*. In particular, if GDF-9 plays a role in oocyte maturation, it may be useful in *in vitro* fertilization procedures, e.g., in enhancing the success rate. Many of the members of the TGF-β family are also important mediators of tissue repair. TGF-β has been shown to have marked effects on the formation of collagen and causes a striking angiogenic response in the newborn mouse (Roberts, *et al.*, *Proc. Natl. Acad. Sci. USA*, 83:4167, 1986). GDF-9 may also have similar activities and may be useful in repair of tissue injury caused by trauma or burns for example.

The term "substantially pure" as used herein refers to GDF-9 which is substantially free of other proteins, lipids, carbohydrates or other materials with which it is naturally associated. One skilled in the art can purify GDF-9 using standard techniques for protein purification. The substantially pure polypeptide will yield a single major band on a non-reducing polyacrylamide gel. The purity of the GDF-9 polypeptide can also be determined by amino-terminal amino acid sequence analysis. GDF-9 polypeptide includes functional fragments of the polypeptide, as long as the activity of GDF-9 remains. Smaller peptides containing the biological activity of GDF-9 are included in the invention.

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The invention provides polynucleotides encoding the GDF-9 protein. These polynucleotides include DNA, cDNA and RNA sequences which encode GDF-9. It is understood that all polynucleotides encoding all or a portion of GDF-9 are also included herein, as long as they encode a polypeptide with GDF-9 activity. Such polynucleotides include naturally occurring, synthetic, and intentionally manipulated polynucleotides. For example, GDF-9 polynucleotide may be subjected to site-directed mutagenesis. The polynucleotide sequence for GDF-9 also includes antisense sequences. The polynucleotides of the invention include sequences that are degenerate as a result of the genetic code. There are 20 natural amino acids, most of which are specified by more than one codon. Therefore, all degenerate nucleotide sequences are included in the invention as long as the amino acid sequence of GDF-9 polypeptide encoded by the nucleotide sequence is functionally unchanged.

Specifically disclosed herein is a cDNA sequence for GDF-9 which is 1712 base pairs in length and contains an open reading frame beginning with a methionine codon at nucleotide 29. The encoded polypeptide is 441 amino acids in length with a molecular weight of about 49.6 kD, as determined by nucleotide sequence analysis. The GDF-9 sequence contains a core of hydrophobic amino acids near the N-terminus, suggestive of a signal sequence for secretion. GDF-9 contains four potential N-glycosylation sites at asparagine residues 163, 229, 258, and 325 and a putative tetrabasic proteolytic processing site (RRRR) at amino acids 303-306. The mature C-terminal fragment of GDF-9 is predicted to be 135 amino acids in length and have an unglycosylated molecular weight of about 15.6 kD, as determined by nucleotide sequence analysis. One skilled in the art can modify, or partially or completely remove the glycosyl groups from the GDF-9 protein using standard techniques. Therefore, the functional protein or fragments thereof of the invention includes glycosylated, partially glycosylated and unglycosylated species of GDF-9.

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The degree of sequence identity of GDF-9 with known TGF- β family members ranges from a minimum of 21% with Mullerian inhibiting substance (MIS) to a maximum of 34% with bone morphogenetic protein-4 (BMP-4). GDF-9 specifically disclosed herein differs from the known family members in its pattern of cysteine residues in the C-terminal region. GDF-9 lacks the fourth cysteine of the seven cysteines present in other family members; in place of cysteine at this position, the GDF-9 sequence contains a serine residue. This GDF-9 does not contain a seventh cysteine residue elsewhere in the C-terminal region.

Minor modifications of the recombinant GDF-9 primary amino acid sequence may result in proteins which have substantially equivalent activity as compared to the GDF-9 polypeptide described herein. Such modifications may be deliberate, as by site-directed mutagenesis, or may be spontaneous. All of the polypeptides produced by these modifications are included herein as long as the biological activity of GDF-9 still exists. Further, deletion of one or more amino acids can also result in a modification of the structure of the resultant molecule without significantly altering its biological activity. This can lead to the development of a smaller active molecule which would have broader utility. For example, one can remove amino or carboxy terminal amino acids which are not required for GDF-9 biological activity.

The nucleotide sequence encoding the GDF-9 polypeptide of the invention includes the disclosed sequence and conservative variations thereof. The term "conservative variation" as used herein denotes the replacement of an amino acid residue by another, biologically similar residue. Examples of conservative variations include the substitution of one hydrophobic residue such as isoleucine, valine, leucine or methionine for another, or the substitution of one polar residue for another, such as the substitution of arginine for lysine, glutamic for aspartic acids, or glutamine for asparagine, and the like. The term

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"conservative variation" also includes the use of a substituted amino acid in place of an unsubstituted parent amino acid provided that antibodies raised to the substituted polypeptide also immunoreact with the unsubstituted polypeptide.

DNA sequences of the invention can be obtained by several methods. For example, the DNA can be isolated using hybridization techniques which are well known in the art. These include, but are not limited to: 1) hybridization of genomic or cDNA libraries with probes to detect homologous nucleotide sequences and 2) antibody screening of expression libraries to detect cloned DNA fragments with shared structural features.

Preferably the GDF-9 polynucleotide of the invention is derived from a mammalian organism, and most preferably from a mouse, rat, or human. Screening procedures which rely on nucleic acid hybridization make it possible to isolate any gene sequence from any organism, provided the appropriate probe is available. Oligonucleotide probes, which correspond to a part of the sequence encoding the protein in question, can be synthesized chemically. This requires that short, oligopeptide stretches of amino acid sequence must be known. The DNA sequence encoding the protein can be deduced from the genetic code, however, the degeneracy of the code must be taken into It is possible to perform a mixed addition reaction when the sequence is degenerate. This includes a heterogeneous mixture of denatured double-stranded DNA. For such screening, hybridization is preferably performed on either single-stranded DNA or denatured double-stranded DNA. Hybridization is particularly useful in the detection of cDNA clones derived from sources where an extremely low amount of mRNA sequences relating to the polypeptide of interest are present. In other words, by using stringent hybridization conditions directed to avoid non-specific binding, it is possible, for example, to allow the autoradiographic visualization of a specific cDNA

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-clone by the hybridization_of_the_target DNA to that single probe in the mixture which is its complete complement (Wallace, et al., Nucl. Acid Res., 9:879, 1981).

The development of specific DNA sequences encoding GDF-9 can also be obtained by: 1) isolation of double-stranded DNA sequences from the genomic DNA; 2) chemical manufacture of a DNA sequence to provide the necessary codons for the polypeptide of interest; and 3) *in vitro* synthesis of a double-stranded DNA sequence by reverse transcription of mRNA isolated from a eukaryotic donor cell. In the latter case, a double-stranded DNA complement of mRNA is eventually formed which is generally referred to as cDNA.

Of the three above-noted methods for developing specific DNA sequences for use in recombinant procedures, the isolation of genomic DNA isolates is the least common. This is especially true when it is desirable to obtain the microbial expression of mammalian polypeptides due to the presence of introns.

The synthesis of DNA sequences is frequently the method of choice when the entire sequence of amino acid residues of the desired polypeptide product is known. When the entire sequence of amino acid residues of the desired polypeptide is not known, the direct synthesis of DNA sequences is not possible and the method of choice is the synthesis of cDNA sequences. Among the standard procedures for isolating cDNA sequences of interest is the formation of plasmid- or phage-carrying cDNA libraries which are derived from reverse transcription of mRNA which is abundant in donor cells that have a high level of genetic expression. When used in combination with polymerase chain reaction technology, even rare expression products can be cloned. In those cases where significant portions of the amino acid sequence of the polypeptide are known, the production of labeled single or double-stranded

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DNA or RNA probe sequences duplicating a sequence putatively present in the target cDNA may be employed in DNA/DNA hybridization procedures which are carried out on cloned copies of the cDNA which have been denatured into a single-stranded form (Jay, et al., Nucl. Acid Res., 11:2325, 1983).

A cDNA expression library, such as lambda gt11, can be screened indirectly for GDF-9 peptides having at least one epitope, using antibodies specific for GDF-9. Such antibodies can be either polyclonally or monoclonally derived and used to detect expression product indicative of the presence of GDF-9 cDNA.

DNA sequences encoding GDF-9 can be expressed *in vitro* by DNA transfer into a suitable host cell. "Host cells" are cells in which a vector can be propagated and its DNA expressed. The term also includes any progeny of the subject host cell. It is understood that all progeny may not be identical to the parental cell since there may be mutations that occur during replication. However, such progeny are included when the term "host cell" is used. Methods of stable transfer, meaning that the foreign DNA is continuously maintained in the host, are known in the art.

In the present invention, the GDF-9 polynucleotide sequences may be inserted into a recombinant expression vector. The term "recombinant expression vector" refers to a plasmid, virus or other vehicle known in the art that has been manipulated by insertion or incorporation of the GDF-9 genetic sequences. Such expression vectors contain a promoter sequence which facilitates the efficient transcription of the inserted genetic sequence of the host. The expression vector typically contains an origin of replication, a promoter, as well as specific genes which allow phenotypic selection of the transformed cells. Vectors suitable for use in the present invention include, but are not limited to the T7-based expression vector for expression in bacteria (Rosenberg, et al.,

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Gene ,56:125, 1987), the pMSXND expression vector for expression in mammalian cells (Lee and Nathans, *J. Biol. Chem.*, 263:3521, 1988) and baculovirus-derived vectors for expression in insect cells. The DNA segment can be present in the vector operably linked to regulatory elements, for example, a promoter (e.g., T7, metallothionein I, or polyhedrin promoters).

Polynucleotide sequences encoding GDF-9 can be expressed in either prokaryotes or eukaryotes. Hosts can include microbial, yeast, insect and mammalian organisms. Methods of expressing DNA sequences having eukaryotic or viral sequences in prokaryotes are well known in the art. Biologically functional viral and plasmid DNA vectors capable of expression and replication in a host are known in the art. Such vectors are used to incorporate DNA sequences of the invention.

Transformation of a host cell with recombinant DNA may be carried out by conventional techniques as are well known to those skilled in the art. Where the host is prokaryotic, such as *E. coli*, competent cells which are capable of DNA uptake can be prepared from cells harvested after exponential growth phase and subsequently treated by the CaCl₂ method using procedures well known in the art. Alternatively, MgCl₂ or RbCl can be used. Transformation can also be performed after forming a protoplast of the host cell if desired.

When the host is a eukaryote, such methods of transfection of DNA as calcium phosphate co-precipitates, conventional mechanical procedures such as microinjection, electroporation, insertion of a plasmid encased in liposomes, or virus vectors may be used. Eukaryotic cells can also be cotransformed with DNA sequences encoding the GDF-9 of the invention, and a second foreign DNA molecule encoding a selectable phenotype, such as the herpes simplex thymidine kinase gene. Another method is to use a eukaryotic viral vector, such as simian virus 40 (SV40) or bovine papilloma virus, to transiently infect

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or transform eukaryotic cells and express the protein. (see for example, Eukaryotic Viral Vectors, Cold Spring Harbor Laboratory, Gluzman ed., 1982).

Isolation and purification of microbial expressed polypeptide, or fragments thereof, provided by the invention, may be carried out by conventional means including preparative chromatography and immunological separations involving monoclonal or polyclonal antibodies.

The invention includes antibodies immunoreactive with GDF-9 polypeptide or functional fragments thereof. Antibody which consists essentially of pooled monoclonal antibodies with different epitopic specificities, as well as distinct monoclonal antibody preparations are provided. Monoclonal antibodies are made from antigen containing fragments of the protein by methods well known to those skilled in the art (Kohler, et al., Nature, 256:495, 1975). The term antibody as used in this invention is meant to include intact molecules as well as fragments thereof, such as Fab and F(ab')₂, which are capable of binding an epitopic determinant on GDF-9.

The term "cell-proliferative disorder" denotes malignant as well as non-malignant cell populations which often appear to differ from the surrounding tissue both morphologically and genotypically. The GDF-9 polynucleotide that is an antisense molecule is useful in treating malignancies of the various organ systems, particularly, for example, the ovaries. Essentially, any disorder which is etiologically linked to altered expression of GDF-9 could be considered susceptible to treatment with a GDF-9 suppressing reagent.

The invention provides a method for detecting a cell proliferative disorder of the ovary which comprises contacting an anti-GDF-9 antibody with a cell suspected of having a GDF-9 associated disorder and detecting binding to the antibody. The antibody reactive with GDF-9 is labeled with a compound which allows

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specific for GDF-9 polypeptide may be used to detect the level of GDF-9 in biological fluids and tissues. Any specimen containing a detectable amount of antigen can be used. A preferred sample of this invention is tissue of ovarian origin, specifically tissue containing granulosa cells or ovarian follicular fluid. The level of GDF-9 in the suspect cell can be compared with the level in a normal cell to determine whether the subject has a GDF-9-associated cell proliferative disorder. Preferably the subject is human.

The antibodies of the invention can be used in any subject in which it is desirable to administer *in vitro* or *in vivo* immunodiagnosis or immunotherapy. The antibodies of the invention are suited for use, for example, in immunoassays in which they can be utilized in liquid phase or bound to a solid phase carrier. In addition, the antibodies in these immunoassays can be detectably labeled in various ways. Examples of types of immunoassays which can utilize antibodies of the invention are competitive and non-competitive immunoassays in either a direct or indirect format. Examples of such immunoassays are the radioimmunoassay (RIA) and the sandwich (immunometric) assay. Detection of the antigens using the antibodies of the invention can be done utilizing immunoassays which are run in either the forward, reverse, or simultaneous modes, including immunohistochemical assays on physiological samples. Those of skill in the art will know, or can readily discern, other immunoassay formats without undue experimentation.

The antibodies of the invention can be bound to many different carriers and used to detect the presence of an antigen comprising the polypeptide of the invention. Examples of well-known carriers include glass, polystyrene. polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, agaroses and magnetite. The nature of the carrier can be either soluble or insoluble for purposes of the invention. Those skilled

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in the art will know of other suitable carriers for binding antibodies, or will be able to ascertain such, using routine experimentation.

There are many different labels and methods of labeling known to those of ordinary skill in the art. Examples of the types of labels which can be used in the present invention include enzymes, radioisotopes, fluorescent compounds, colloidal metals, chemiluminescent compounds, phosphorescent compounds, and bioluminescent compounds. Those of ordinary skill in the art will know of other suitable labels for binding to the antibody, or will be able to ascertain such, using routine experimentation.

Another technique which may also result in greater sensitivity consists of coupling the antibodies to low molecular weight haptens. These haptens can then be specifically detected by means of a second reaction. For example, it is common to use such haptens as biotin, which reacts with avidin, or dinitrophenyl, puridoxal, and fluorescein, which can react with specific anti-hapten antibodies.

In using the monoclonal antibodies of the invention for the *in vivo* detection of antigen, the detectably labeled antibody is given a dose which is diagnostically effective. The term "diagnostically effective" means that the amount of detectably labeled monoclonal antibody is administered in sufficient quantity to enable detection of the site having the antigen comprising a polypeptide of the invention for which the monoclonal antibodies are specific.

The concentration of detectably labeled monoclonal antibody which is adminstered should be sufficient such that the binding to those cells having the polypeptide is detectable compared to the background. Further, it is desirable that the detectably labeled monoclonal antibody be rapidly cleared from the circulatory system in order to give the best target-to-background signal ratio.

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As a rule, the dosage of detectably labeled monoclonal antibody for in vivo diagnosis will vary depending on such factors as age, sex, and extent of disease of the individual. Such dosages may vary, for example, depending on whether multiple injections are given, antigenic burden, and other factors known to those of skill in the art.

For *in vivo* diagnostic imaging, the type of detection instrument available is a major factor in selecting a given radioisotope. The radioisotope chosen must have a type of decay which is detectable for a given type of instrument. Still another important factor in selecting a radioisotope for *in vivo* diagnosis is that deleterious radiation with respect to the host is minimized. Ideally, a radioisotope used for *in vivo* imaging will lack a particle emission, but produce a large number of photons in the 140-250 keV range, which may readily be detected by conventional gamma cameras.

For *in vivo* diagnosis radioisotopes may be bound to immunoglobulin either directly or indirectly by using an intermediate functional group. Intermediate functional groups which often are used to bind radioisotopes which exist as metallic ions to immunoglobulins are the bifunctional chelating agents such as diethylenetriaminepentacetic acid (DTPA) and ethylenediaminetetraacetic acid (EDTA) and similar molecules. Typical examples of metallic ions which can be bound to the monoclonal antibodies of the invention are ¹¹¹In, ⁹⁷Ru, ⁶⁷Ga, ⁶⁸Ga, ⁷²As, ⁸⁹Zr, and ²⁰¹Tl.

The monoclonal antibodies of the invention can also be labeled with a paramagnetic isotope for purposes of *in vivo* diagnosis, as in magnetic resonance imaging (MRI) or electron spin resonance (ESR). In general, any conventional method for visualizing diagnostic imaging can be utilized. Usually gamma and positron emitting radioisotopes are used for camera imaging and

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paramagnetic isotopes for MRI. Elements which are particularly useful in such techniques include ¹⁵⁷Gd, ⁵⁵Mn, ¹⁶²Dy, ⁵²Cr, and ⁵⁶Fe.

The monoclonal antibodies of the invention can be used *in vitro* and *in vivo* to monitor the course of amelioration of a GDF-9-associated disease in a subject. Thus, for example, by measuring the increase or decrease in the number of cells expressing antigen comprising a polypeptide of the invention or changes in the concentration of such antigen present in various body fluids, it would be possible to determine whether a particular therapeutic regimen aimed at ameliorating the GDF-9-associated disease is effective. The term "ameliorate" denotes a lessening of the detrimental effect of the GDF-9-associated disease in the subject receiving therapy.

The present invention identifies a nucleotide sequence that can be expressed in an altered manner as compared to expression in a normal cell, therefore, it is possible to design appropriate therapeutic or diagnostic techniques directed to this sequence. Thus, where a cell-proliferative disorder is associated with the expression of GDF-9, nucleic acid sequences that interfere with GDF-9 expression at the translational level can be used. This approach utilizes, for example, antisense nucleic acid and ribozymes to block translation of a specific GDF-9 mRNA, either by masking that mRNA with an antisense nucleic acid or by cleaving it with a ribozyme.

Antisense nucleic acids are DNA or RNA molecules that are complementary to at least a portion of a specific mRNA molecule (Weintraub, Scientific American, 262:40, 1990). In the cell, the antisense nucleic acids hybridize to the corresponding mRNA, forming a double-stranded molecule. The antisense nucleic acids interfere with the translation of the mRNA, since the cell will not translate a mRNA that is double-stranded. Antisense oligomers of about 15 nucleotides are preferred, since they are easily synthesized and are less likely

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to cause problems than larger molecules when introduced into the target GDF-9-producing cell. The use of antisense methods to inhibit the *in vitro* translation of genes is well known in the art (Marcus-Sakura, *Anal.Biochem.*, 172:289, 1988).

other single-stranded RNA in a manner analogous to DNA restriction endonucleases. Through the modification of nucleotide sequences which encode these RNAs, it is possible to engineer molecules that recognize specific nucleotide sequences in an RNA molecule and cleave it (Cech, *J.Amer.Med. Assn.*, 260:3030, 1988). A major advantage of this approach is that, because they are sequence-specific, only mRNAs with particular sequences are inactivated.

There are two basic types of ribozymes namely, tetrahymena-type (Hasselhoff, Nature, 334:585, 1988) and "hammerhead"-type. Tetrahymena-type ribozymes recognize sequences which are four bases in length, while "hammerhead"-type ribozymes recognize base sequences 11-18 bases in length. The longer the recognition sequence, the greater the likelihood that the sequence will occur exclusively in the target mRNA species. Consequently, hammerhead-type ribozymes are preferable to tetrahymena-type ribozymes for inactivating a specific mRNA species and 18-based recognition sequences are preferable to shorter recognition sequences.

The present invention also provides gene therapy for the treatment of cell proliferative disorders which are mediated by GDF-9 protein. Such therapy would achieve its therapeutic effect by introduction of the GDF-9 antisense polynucleotide into cells having the proliferative disorder. Delivery of antisense GDF-9 polynucleotide can be achieved using a recombinant expression vector such as a chimeric virus or a colloidal dispersion system.

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Especially preferred for therapeutic delivery of antisense sequences is the use of targeted liposomes.

Various viral vectors which can be utilized for gene therapy as taught herein include adenovirus, herpes virus, vaccinia, or, preferably, an RNA virus such as a retrovirus. Preferably, the retroviral vector is a derivative of a murine or avian retrovirus. Examples of retroviral vectors in which a single foreign gene can be inserted include, but are not limited to: Moloney murine leukemia virus (MoMuLV), Harvey murine sarcoma virus (HaMuSV), murine mammary tumor virus (MuMTV), and Rous Sarcoma Virus (RSV). A number of additional retroviral vectors can incorporate multiple genes. All of these vectors can transfer or incorporate a gene for a selectable marker so that transduced cells can be identified and generated. By inserting a GDF-9 sequence of interest into the viral vector, along with another gene which encodes the ligand for a receptor on a specific target cell, for example, the vector is now target specific. Retroviral vectors can be made target specific by inserting, for example, a polynucleotide encoding a sugar, a glycolipid, or a protein. Preferred targeting is accomplished by using an antibody to target the retroviral vector. Those of skill in the art will know of, or can readily ascertain without undue experimentation, specific polynucleotide sequences which can be inserted into the retroviral genome to allow target specific delivery of the retroviral vector containing the GDF-9 antisense polynucleotide.

Since recombinant retroviruses are defective, they require assistance in order to produce infectious vector particles. This assistance can be provided, for example, by using helper cell lines that contain plasmids encoding all of the structural genes of the retrovirus under the control of regulatory sequences within the LTR. These plasmids are missing a nucleotide sequence which enables the packaging mechanism to recognize an RNA transcript for encapsidation. Helper cell lines which have deletions of the packaging signal

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include, but are not-limited to \$\psi_2\$, PA317 and PA12, for example. These cell lines produce empty virions, since no genome is packaged. If a retroviral vector is introduced into such cells in which the packaging signal is intact, but the structural genes are replaced by other genes of interest, the vector can be packaged and vector virion produced.

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Alternatively, NIH 3T3 or other tissue culture cells can be directly transfected with plasmids encoding the retroviral structural genes *gag*, *pol* and *env*, by conventional calcium phosphate transfection. These cells are then transfected with the vector plasmid containing the genes of interest. The resulting cells release the retroviral vector into the culture medium.

Another targeted delivery system for GDF-9 antisense polynucleotides is a colloidal dispersion system. Colloidal dispersion systems include macromolecule complexes, nanocapsules, microspheres, beads, and lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. The preferred colloidal system of this invention is a liposome. Liposomes are artificial membrane vesicles which are useful as delivery vehicles in vitro and in vivo. It has been shown that large unilamellar vesicles (LUV), which range in size from 0.2-4.0 µm can encapsulate a substantial percentage of an aqueous buffer containing large macromolecules. RNA, DNA and intact virions can be encapsulated within the aqueous interior and be delivered to cells in a biologically active form (Fraley, et al., Trends Biochem. Sci., 6:77, 1981). In addition to mammalian cells, liposomes have been used for delivery of polynucleotides in plant, yeast and bacterial cells. In order for a liposome to be an efficient gene transfer vehicle, the following characteristics should be present; (1) encapsulation of the genes of interest at high efficiency while not compromising their biological activity; (2) preferential and substantial binding to a target cell in comparison to non-target cells; (3) delivery of the aqueous contents of the vesicle to the target cell cytoplasm at high efficiency; and (4)

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accurate and effective expression of genetic information (Mannino, et al., Biotechniques, 6:682, 1988).

The composition of the liposome is usually a combination of phospholipids, particularly high-phase-transition-temperature phospholipids, usually in combination with steroids, especially cholesterol. Other phospholipids or other lipids may also be used. The physical characteristics of liposomes depend on pH, ionic strength, and the presence of divalent cations.

Examples of lipids useful in liposome production include phosphatidyl compounds, such as phosphatidylglycerol, phosphatidylcholine, phosphatidylserine, phosphatidylethanolamine, sphingolipids, cerebrosides, and gangliosides. Particularly useful are diacylphosphatidylglycerols, where the lipid moiety contains from 14-18 carbon atoms, particularly from 16-18 carbon atoms, and is saturated. Illustrative phospholipids include egg phosphatidylcholine, dipalmitoylphosphatidylcholine and distearoylphosphatidylcholine.

The targeting of liposomes can be classified based on anatomical and mechanistic factors. Anatomical classification is based on the level of selectivity, for example, organ-specific, cell-specific, and organelle-specific. Mechanistic targeting can be distinguished based upon whether it is passive or active. Passive targeting utilizes the natural tendency of liposomes to distribute to cells of the reticulo-endothelial system (RES) in organs which contain sinusoidal capillaries. Active targeting, on the other hand, involves alteration of the liposome by coupling the liposome to a specific ligand such as a monoclonal antibody, sugar, glycolipid, or protein, or by changing the composition or size of the liposome in order to achieve targeting to organs and cell types other than the naturally occurring sites of localization.

The surface of the targeted delivery system may be modified in a variety of ways. In the case of a liposomal targeted delivery system, lipid groups can be incorporated into the lipid bilayer of the liposome in order to maintain the targeting ligand in stable association with the liposomal bilayer. Various linking groups can be used for joining the lipid chains to the targeting ligand.

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Due to the expression of GDF-9 in the reproductive tract, there are a variety of applications using the polypeptide, polynucleotide and antibodies of the invention, related to contraception, fertility and pregnancy. GDF-9 could play a role in regulation of the menstrual cycle and, therefore, could be useful in various contraceptive regimens.

The following examples are intended to illustrate but not limit the invention. While they are typical of those that might be used, other procedures known to those skilled in the art may alternatively be used.

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EXAMPLE 1 IDENTIFICATION AND ISOLATION OF A NOVEL TGF-B FAMILY MEMBER

To identify a new member of the TGF- β superfamily, degenerate oligonucleotides were designed which corresponded to two conserved regions among the known family members: one region spanning the two tryptophan residues conserved in all family members except MIS and the other region spanning the invariant cysteine residues near the C-terminus. These primers were used for polymerase chain reactions on mouse genomic DNA followed by subcloning the PCR products using restriction sites placed at the 5' ends of the primers, picking individual *E. coli* colonies carrying these subcloned inserts, and using a combination of random sequencing and hybridization analysis to eliminate known members of the superfamily.

GDF-9 was identified from a mixture of PCR products obtained with the primers SJL160 (5'-CCGGAATTCGGITGG(G/C/A)A(G/A/T/C)(G/C/A)A(G/A/T/C) TGG(A/G)TI(A/G)TI(T/G)CICC-3') (SEQUENCE ID NO. 1) and SJL153 (5'-C C G G A A T T C (A / G) C A I (G / C) C (A / G) C A I C (T / C) (G / A / T - /C)(C/G/T)TIG(T/C)I(G/A)(T/C)CAT-3') (SEQUENCE ID NO. 2). PCR using these primers was carried out with 2 μ g mouse genomic DNA at 94°C for 1 min, 50°C for 2 min, and 72°C for 2 min for 40 cycles.

PCR products of approximately 280 bp were gel-purified, digested with Eco RI, gel-purified again, and subcloned in the Bluescript vector (Stratagene, San Diego, CA). Bacterial colonies carrying individual subclones were picked into 96 well microtiter plates, and multiple replicas were prepared by plating the cells onto nitrocellulose. The replicate filters were hybridized to probes representing known members of the family, and DNA was prepared from non-hybridizing colonies for sequence analysis.

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The primer combination—of SJL160 and SJL153, yielded three known sequences (inhibin β B, BMP-2, and BMP-4) and one novel sequence (designated GDF-9) among 145 subclones analyzed.

RNA isolation and Northern analysis were carried out as described previously (Lee,S.J., *Mol. Endocrinol.* 4:1034, 1990). An oligo dT-primed cDNA library was prepared from 2.5-3 μ g of ovary poly A-selected RNA in the lambda ZAP II vector according to the instructions provided by Stratagene. The ovary library was not amplified prior to screening. Filters were hybridized as described previously (Lee, S.-J., *Proc. Natl. Acad. Sci. USA.*, 88:4250-4254, 1991). DNA sequencing of both strands was carried out using the dideoxy chain termination method (Sanger, *et al., Proc. Natl. Acad. Sci., USA*, 74:5463-5467, 1977) and a combination of the S1 nuclease/exonuclease III strategy (Henikoff, S., *Gene*, 28:351-359, 1984) and synthetic oligonucleotide primers.

EXAMPLE 2 EXPRESSION PATTERN AND SEQUENCE OF GDF-9

To determine the expression pattern of GDF-9, RNA samples prepared from a variety of adult tissues were screened by Northern analysis. Five micrograms of twice polyA-selected RNA prepared from each tissue were electrophoresed on formaldehyde gels, blotted and probed with GDF-9. As shown in Figure 1, the GDF-9 probe detected a 1.7 kb mRNA expressed exclusively in the ovary.

A mouse ovary cDNA library of 1.5 x 10⁶ recombinant phage was constructed in lambda ZAP II and screened with a probe derived from the GDF-9 PCR product. The nucleotide sequence of the longest of nineteen hybridizing clones is shown in Figure 2. Consensus N-glycosylation signals are denoted by plain boxes. The putative tetrabasic processing sites are denoted by

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stippled boxes. The in-frame-termination codons upstream of the putative initiating ATG and the consensus polyadenylation signals are underlined. The poly A tails are not shown. Numbers indicate nucleotide position relative to the 5' end. The 1712 bp sequence contains a long open reading frame beginning with a methionine codon at nucleotide 29 and potentially encoding a protein 441 amino acids in length with a molecular weight of 49.6 kD. Like other TGF- ρ family members, the GDF-9 sequence contains a core of hydrophobic amino acids near the N-terminus suggestive of a signal sequence for secretion. GDF-9 contains four potential N-glycosylation sites at asparagine residues 163, 229, 258, and 325 and a putative tetrabasic proteolytic processing site (RRRR) at amino acids 303-306. The mature C-terminal fragment of GDF-9 is predicted to be 135 amino acids in length and have an unglycosylated molecular weight of 15.6 kD.

Although the C-terminal portion of GDF-9 clearly shows homology with the other family members, the sequence of GDF-9 is significantly diverged from those of the other family members (Figures 3 and 4). Figure 3 shows the alignment of the C-terminal sequences of GDF-9 with the corresponding regions of human GDF-1 (Lee, Proc. Natl. Acad. Sci. USA, 88:4250-4254, 1991), Xenopus Vg-1 (Weeks, et al., Cell, 51:861-867, 1987), human Vgr-1 (Celeste, et al., Proc. Natl. Acad. Sci. USA, 87:9843-9847, 1990), human OP-1 (Ozkaynak, et al., EMBO J., 9:2085-2093, 1990), human BMP-5 (Celeste, et al., Proc. Natl. Acad. Sci. USA, 87:9843-9847, 1990), Drosophila 60A (Wharton, et al., Proc. Natl. Acad. Sci. USA, 88:9214-9218, 1991), human BMP-2 and 4 (Wozney, et al., Science, 242:1528-1534, 1988), Drosophila DPP (Padgett, et al., Nature, 325:81-84, 1987), human BMP-3 (Wozney, et al., Science, 242:1528-1534, 1988), human MIS (Cate, et al., Cell, 45:685-698, 1986), human inhibin, β A, and β B (Mason, et al., Biochem, Biophys. Res. Commun., 135:957-964, 1986), human TGF-β1 (Derynck, et al., Nature, 316:701-705, 1985), humanTGF-β2 (deMartin, et al., EMBO J., 6:3673-3677, 1987), human TGF-β3

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(ten Dijke, et al.,-Proc. Natl. Acad. Sci. USA, 85:4715-4719, 1988), chicken TGFβ4 (Jakowlew, et al., Mol. Endocrinol., 2:1186-1195, 1988), and Xenopus TGFβ5 (Kondaiah, et al., J. Biol. Chem., 265:1089-1093, 1990). The conserved cysteine residues are shaded. Dashes denote gaps introduced in order to maximize the alignment.

Figure 4 shows the amino acid homologies among the different members of the TGF- ρ superfamily. Numbers represent percent amino acid identities between each pair calculated from the first conserved cysteine to the C-terminus. Boxes represent homologies among highly-related members within particular subgroups.

The degree of sequence identify with known family members ranges from a minimum of 21% with MIS to a maximum of 34% with BMP-4. Hence, GDF-9 is comparable to MIS in its degree of sequence divergence from the other members of this superfamily. Moreover, GDF-9 shows no significant sequence homology to other family members in the pro-region of the molecule. GDF-9 also differs from the known family members in its pattern of cysteine residues in the C-terminal region. GDF-9 lacks the fourth cysteine of the seven cysteines that are present in all other family members; in place of cysteine at this position, the GDF-9 sequence contains a serine residue. In addition, GDF-9 does not contain a seventh cysteine residue elsewhere in the C-terminal region.

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IMMUNOCHEMICAL LOCALIZATION OF GDF-9 IN THE ZONA PELLUCIDA

To determine whether GDF-9 mRNA was translated, sections of adult ovaries were incubated with antibodies directed against recombinant GDF-9 protein. In order to raise antibodies against GDF-9, portions of GDF-9 cDNA spanning amino acids 30 to 295 (pro-region) or 308 to 441 (mature region) were cloned into the T7-based pET3 expression vector (provided by F.W. Studier, Brookhaven National Laboratory), and the resulting plasmids were transformed into the BL21 (DE3) bacterial strain. Total cell extracts from isopropyl B-D-thiogalactoside-induced cells were electrophoresed on SDS/polyacrylamide gels, and the GDF-9 protein fragments were excised, mixed with Freund's adjuvant, and used to immunize rabbits by standard methods known to those of skill in the art. All immunizations were carried out by Spring Valley Lab (Sykesville, MD). The presence of GDF-9-reactive antibodies in the sera of these rabbits was assessed by Western analysis of bacterially-expressed protein fragments. The resulting serum was shown to react with the bacterially-expressed protein by Western analysis.

For immunohistochemical studies, ovaries were removed from adult mice, fixed in 4% paraformaldehyde, embedded in paraffin, and sectioned. Sites of antibody binding were detected by using the Vectastain ABC kit, according to the instructions provided by Vector Laboratories. FIGURE 5 shows the immunohistochemical localization of GDF-9 protein. Adjacent sections of an adult ovary were either stained with hematoxylin and eosin (FIGURE 5a) or incubated with immune (FIGURE 5b) or pre-immune (FIGURE 5c) serum at a dilution of 1:500. As shown in FIGURE 5b, the antiserum detected protein

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solely in oocytes. No staining was detected using pre-immune serum (FIGURE 5c). Hence, GDF-9 protein appears to translated *in vivo* by oocytes.

EXAMPLE 4 ISOLATION OF HUMAN GDF-9

In order to isolate a cDNA clone encoding human GDF-9, a cDNA library was constructed in lambda ZAP II using poly A-selected RNA prepared from an adult human ovary. From this library, a cDNA clone containing the entire human GDF-9 coding sequence was identified using standard screening techniques as in Example 1 and using the murine GDF-9 clone as a probe. A comparison of the predicted amino acid sequences of murine (top lines) and human (bottom lines) GDF-9 is shown in FIGURE 6. Numbers represent amino acid positions relative to the N-termini. Vertical lines represent sequence identities. Dots represent gaps introduced in order to maximize the alignment. The clear box shows the predicted proteolytic processing sites. The shaded boxes show the cysteine residues in the mature region of the proteins. The bars at the bottom show a schematic of the pre-(clear) and mature (shaded) regions of GDF-9 with the percent sequence identities between the murine and human sequences shown below.

Like murine GDF-9, human GDF-9 contains a hydrophobic leader sequence, a putative RXXR proteolytic cleavage site, and a C-terminal region containing the hallmarks of other TGF- ρ family members. Murine and human GDF-9 are 64% identical in the pro- region and 90% identical in the predicted mature region of the molecule. The high degree of homology between the two sequences suggests that human GDF-9 plays an important role during embryonic development and/or in the adult ovary.

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EXAMPLE 5

NUCLEIC ACID DETECTION OF EXPRESSION OF GDF-9 IN OOCYTES

In order to localize the expression of GDF-9 in the ovary, *in situ* hybridization to mouse ovary sections was carried out using an antisense GDF-9 RNA probe. FIGURE 7 shows *in situ* hybridization to adult ovary sections using a GDF-9 RNA probe. [³⁵S]-labeled anti-sense (FIGURE 7a and 7c) or sense (FIGURE 7 b and 7d) GDF-9 RNA probes were hybridized to adjacent paraffinembedded sections of ovaries fixed in 4% paraformaldehyde. Sections were dipped in photographic emulsion, exposed, developed, and then stained with hematoxylin and eosin. Two representative fields are shown.

As shown in FIGURES 7a and 7c, GDF-9 mRNA was detected primarily in occytes in adult ovaries. Every occyte (regardless of the stage of follicular development) examined showed GDF-9 expression, and no expression was detected in any other cell types. No hybridization was seen using a control GDF-9 sense RNA probe (FIGURE 7b and 7d). Hence, GDF-9 expression appears to be occyte-specific in adult ovaries.

To determine the pattern of expression of GDF-9 mRNA during ovarian development, sections of neonatal ovaries were probed with a GDF-9 RNA probe. FIGURE 8 shows *in situ* hybridization to a postnatal day 4 ovary section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 8a) or dark-field (FIGURE 8b) illumination.

FIGURE 9 shows *in situ* hybridization to postnatal day 8 ovary sections using an antisense (FIGURE 9a) or sense (FIGURE 9b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

GDF-9 mRNA expression was first detected at the onset of follicular development. This was most clearly evident at postnatal day 4, where only occytes that were present in follicles showed GDF-9 expression (FIGURE 8); no expression was seen in occytes that were not surrounded by granulosa cells. By postnatal day 8, every occyte appeared to have undergone follicular development, and every occyte showed GDF-9 expression (FIGURE 9).

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To determine whether GDF-9 was also expressed following ovulation, sections of mouse oviducts were examined by *in situ* hybridization. FIGURE 10 shows *in situ* hybridization to adult oviduct sections using an antisense (FIGURE 10a) or sense (FIGURE 10b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

FIGURE 11 shows *in situ* hybridization to an adult oviduct (0.5 days following fertilization) section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 11a) or dark-field (FIGURE 11b) illumination.

As shown in FIGURE 10, GDF-9 was expressed by oocytes that had been released into the oviduct. However, the expression of GDF-9 mRNA turned off rapidly following fertilization of the oocytes; by day 0.5 following fertilization, only some embryos (such as the one shown in FIGURE 11) expressed GDF-9 mRNA, and by day 1.5, all embryos were negative for GDF-9 expression.

Although the invention has been described with reference to the presently preferred embodiment, it should be understood that various modifications can be made without departing from the spirit of the invention. Accordingly, the invention is limited only by the following claims.

SUMMARY OF SEQUENCES

Sequence ID No. 1 is the nucleotide sequence for the primer, SJL160, for GDF-9 (page 24, lines 15 and 16);

Sequence ID No. 2 is the nucleotide sequence for the primer, SJL153, for GDF-9 (page 24, lines 17 and 18);

Sequence ID No. 3 is the nucleotide and deduced amino acid sequence for GDF-9 (Figure 2);

Sequence ID No. 4 is the deduced amino acid sequence for GDF-9 (Figure 2);

Sequence ID No. 5 is the amino acid sequence of the C-terminus of GDF-3 (Figure 3);

Sequence ID No. 6 is the amino acid sequence of the C-terminus of GDF-9 (Figure 3);

Sequence ID No. 7 is the amino acid sequence of the C-terminus of GDF-1 (Figure 3);

Sequence ID No. 8 is the amino acid sequence of the C-terminus of Vg-1 (Figure 3);

Sequence ID No. 9 is the amino acid sequence of the C-terminus of Vgr-1 (Figure 3);

Sequence ID No. 10 is the amino acid sequence of the C-terminus of OP-1 (Figure 3);

Sequence ID No. 11 is the amino acid sequence of the C-terminus of BMP-5 (Figure 3);

Sequence ID No. 12 is the amino acid sequence of the C-terminus of 60A (Figure 3);

Sequence ID No. 13 is the amino acid sequence of the C-terminus of BMP-2 (Figure 3);

Sequence ID No. 14 is the amino acid sequence of the C-terminus of BMP-4 (Figure 3);

Sequence ID No. 15 is the amino acid sequence of the C-terminus of DPP (Figure 3);

Sequence ID No. 16 is the amino acid sequence of the C-terminus of BMP-3 (Figure 3);

Sequence ID No. 17 is the amino acid sequence of the C-terminus of MIS (Figure 3);

Sequence ID No. 18 is the amino acid sequence of the C-terminus of inhibin α (Figure 3);

Sequence ID No. 19 is the amino acid sequence of the C-terminus of inhibin β A (Figure 3);

Sequence ID No. 20 is the amino acid sequence of the C-terminus of inhibin βB (Figure 3);

Sequence ID No. 21 is the amino acid sequence of the C-terminus of TGF-\$1 (Figure 3);

Sequence ID No. 22 is the amino acid sequence of the C-terminus of TGF- β 2 (Figure 3);

5 Sequence ID No. 23 is the amino acid sequence of the C-terminus of TGF-β3 (Figure 3);

Sequence ID No. 24 is the amino acid sequence of the C-terminus of TGF- β 4 (Figure 3);

Sequence ID No. 25 is the amino acid sequence of the C-terminus of TGF-\$5 (Figure 3); and

Sequence ID No. 26 is the amino acid sequence of human GDF-9 (Figure 6).

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SEQUENCE LISTING _

(1) GENERAL INFORMATI	ON:
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- (i) APPLICANT: THE JOHNS HOPKINS UNIVERSITY
- (ii) TITLE OF INVENTION: GROWTH DIFFERENTIATION FACTOR-9
- 5 (iii) NUMBER OF SEQUENCES: 26
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Spensley Horn Jubas & Lubitz
 - (B) STREET: 1880 Century Park East, Suite 500
- 10 (C) CITY: Los Angeles

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- (D) STATE: California
- (E) COUNTRY: US
- (F) ZIP: 90067
- (v) COMPUTER READABLE FORM:
- (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
- 20 (A) APPLICATION NUMBER:
 - (B) FILING DATE: 12-JAN-1994
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Wetherell, Jr. Ph.D., John R.
 - (B) REGISTRATION NUMBER: 31,678
 - (C) REFERENCE/DOCKET NUMBER: FD3288
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (619) 455-5100
 - (B) TELEFAX: (619) 455-5110
- 30 (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 35 base pairs
 - (B) TYPE: nucleic acid

(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(11) MOLECULE TYPE: DNA (genomic)	
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,	
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inosine"	
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(2) INFORMATION FOR SEQ ID NO:2:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 33 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(vii) IMMEDIATE SOURCE: (B) CLONE: SJL153	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 133	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:	
CCGGAATTCR CADSCRCADC YNBTDGYDRY CAT	33
(2) INFORMATION FOR SEQ ID NO:3:	
	(D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (vii) IMMEDIATE SOURCE:

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	(vii)	IMM		TE S ONE:												
10		(ix)		A) NA	: ME/K DCATI			1351	Ĺ					٠			
		(xi)	SEC	QUENC	E DE	ESCRI	PTIC	ON: S	SEQ I	D NO):3:						
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			GTT Val													٠	100
20			GCT														148
			GAG Glu														196
25			TCT Ser		Leu										AGG Arg		244
30				Thr					Pro						TAC Tyr		292

									AAA Lys					340
5									CGG Arg 115					388
									GTG Val					436
10									GTG Val	_				484
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	 	Pro				Tyr				Ser			GCC Ala	820

-41-

	TAC Tyr 265														868	
5	CCC Pro														916	
											AAA Lys				964	
10											AAC Asn 325				1012	1
15											GAA Glu				1060)
											TGG Trp				1108	}
20.					Pro						TGT Cys				1156	5
				Tyr					His				Asn	ATA Ile	1204	4
25			Glu	Leu				Val				Cys		CCG Pro	125	2
30	Gly		Tyr				Va]				ı Pro			TCC Ser	130	0
		Ala				Glu				Th:				Cys 440	134	8

	CGT TAGCATGGGG GCCACTTCAA CAAGCCTGCC TGGCAGAGCA ATGCTGTGGG Arg	1401
	CCTTAGAGTG CCTGGGCAGA GAGCTTCCTG TGACCAGTCT CTCCGTGCTG CTCAGTGCAC	1461
5	ACTGTGTGAG CGGGGGAAGT GTGTGTGTGT GGATGAGCAC ATCGAGTGCA GTGTCCGTAG	1521
	GTGTAAAGGG CACACTCACT GGTCGTTGCC ATAAACCAAG TGAAATGTAA CTCATTTGGA	1581
	GAGCTCTTTC TCCCCACGAG TGTAGTTTTC AGTGGACAGA TTTGTTAGCA TAAGTCTCGA	1641
	GTAGAATGTA GCTGTGAACA TGTCAGAGTG CTGTGGTTTT ATGTGACGGA AGAATAAACT	1701
	GTTGATGGCA T	1712
. 10	(2) INFORMATION FOR SEQ ID NO:4:	
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 441 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
15	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
	Met Ala Leu Pro Ser Asn Phe Leu Leu Gly Val Cys Cys Phe Ala Trp 1 5 10 15	
20	Leu Cys Phe Leu Ser Ser Leu Ser Ser Gln Ala Ser Thr Glu Glu Ser 20 25 30	
	Gln Ser Gly Ala Ser Glu Asn Val Glu Ser Glu Ala Asp Pro Trp Ser 35 40 45	
	Leu Leu Pro Val Asp Gly Thr Asp Arg Ser Gly Leu Leu Pro Pro 50 55 60	
25	Leu Phe Lys Val Leu Ser Asp Arg Arg Gly Glu Thr Pro Lys Leu Gln 65 70 75 80	
	Pro Asp Ser Arg Ala Leu Tyr Tyr Met Lys Lys Leu Tyr Lys Thr Tyr 85 90 95	

	Ala	inr	Lys	100	GIY	vai	PIO	Lys	105	ser	ALE	ser	nis	110	lyt	ASII
	Thr	Val	Arg 115	Leu	Phe	Ser	Pro	Cys 120	Ala	Gln	Gln	Glu	Gln 125	Ala	Pro	Ser
5	Asn	Gln 130	Val	Thr	Gly	Pro	Leu 135	Pro	Met	Val	Asp	Leu 140	Leu	Phe	Asn	Leu
	Asp 145	Arg	Val	Thr	Ala	Met 150	Glu	His	Leu	Leu	Lys 155	Ser	Val	Leu	Leu	Tyr 160
10	Thr	Leu	Asn	Asn	Ser 165	Ala	Ser	Ser	Ser	Ser 170	Thr	Val	Thr	Cys	Met 175	Cys
	Asp	Leu	Val	Val 180	Lys	Glu	Ala	Met	Ser 185	Ser	Gly	Arg	Ala	Pro 190	Pro	Arg
	Ala	Pro	Tyr 195	Ser	Phe	Thr	Leu	Lys 200	Lys	His	Arg	Trp	Ile 205	Glu	Ile	Asp
15	Val	Thr 210		Leu	Leu	Gln	Pro 215	Leu	Val	Thr	Ser	Ser 220	Glu	Arg	Ser	Ile
	His 225		Ser	Val	Asn	Phe 230	Thr	Cys	Thr	Lys	Asp 235	Gln	Val	Pro	Glu	Asp 240
20	Gly	Val	Phe	Ser	Met 245	Pro	Leu	Ser	Val	Pro 250	Pro	Ser	Leu	Ile	Leu 255	Tyr
	Leu	Asn	Asp	Thr 260		Thr	Gln	Ala	Tyr 265	His	Ser	Trp	Gln	Ser 270	Leu	G1n
	Ser	Thr	Trp 275	-	Pro	Leu	Gln	His 280		G1y	G1n	Ala	Gly 285	Val	Ala	Ala
25	Arg	Pro 290		Lys	Glu	Glu	Ala 295		Glu	Val	Glu	Arg 300		Pro	Arg	Arg
	Arg 305	_	Gly	Gln	Lys	Ala 310		Arg	Ser	Glu	Ala 315		Gly	Pro	Leu	Leu 320
30	Thr	Ala	. Ser	Phe	Asn 325		Ser	Glu	Tyr	Phe 330		Gln	Phe	Leu	Phe 335	

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Gln Asn Glu Cys Glu Leu His Asp Phe Arg Leu Ser Phe Ser Gln Leu 345 340 Lys Trp Asp Asn Trp Ile Val Ala Pro His Arg Tyr Asn Pro Arg Tyr 360 355 5 · Cys Lys Gly Asp Cys Pro Arg Ala Val Arg His Arg Tyr Gly Ser Pro 370 375 380 Val His Thr Met Val Gln Asn Ile Ile Tyr Glu Lys Leu Asp Pro Ser 390 395 Val Pro Arg Pro Ser Cys Val Pro Gly Lys Tyr Ser Pro Leu Ser Val 10 410 405 Leu Thr Ile Glu Pro Asp Gly Ser Ile Ala Tyr Lys Glu Tyr Glu Asp 425 Met Ile Ala Thr Arg Cys Thr Cys Arg 435 15 (2) INFORMATION FOR SEQ ID NO:5: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 117 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: GDF-3 (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..117 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5: Lys Arg Arg Ala Ala Ile Ser Val Pro Lys Gly Phe Cys Arg Asn Phe 5 10

Cys His Arg His Gln Leu Phe Ile Asn Phe Gln Asp Leu Gly Trp His 30 20 Lys Trp Val Ile Ala Pro Lys Gly Phe Met Ala Asn Tyr Cys His Gly 45 35 40 Glu Cys Pro Phe Ser Met Thr Thr Tyr Leu Asn Ser Ser Asn Tyr Ala 5 60 55 50 Phe Met Gln Ala Leu Met His Met Ala Asp Pro Lys Val Pro Lys Ala 75 70 Val Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Met Leu Tyr Gln Asp 85 90 10 Ser Asp Lys Asn Val Ile Leu Arg His Tyr Glu Asp Met Val Val Asp 105 Glu Cys Gly Cys Gly 115 (2) INFORMATION FOR SEQ ID NO:6: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: GDF-9 (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: Phe Asn Leu Ser Glu Tyr Phe Lys Gln Phe Leu Phe Pro Gln Asn Glu 10 5 1 .

	Cys	Glu	Leu	His 20	Asp	Phe	Arg	Leu	Ser 25	Phe	Ser	Gln	Leu	Lys 30	Trp	Asp
	Asn	Trp	Ile 35	Val	Ala	Pro	His	Arg 40	Tyr	Asn	Pro	-	Tyr 45	Cys	Lys	G1y
5	Asp	Cys 50	Pro	Arg	Ala	Val	Arg 55	His	Arg	Tyr	G1y	Ser 60	Pro	Val	His	Thr
	Met 65	Val	Gln	Asn	Ile	Ile 70	Tyr	G1u	Lys	Leu	Asp 75	Pro	Ser	Val	Pro	Arg 80
10	Pro	Ser	Cys	Val	Pro 85	Gly	Lys	Tyr	Ser	Pro 90	Leu	Ser	Val	Leu	Thr 95	Ile
	Glu	Pro	Asp	Gly 100	Ser	Ile	Ala	Tyr	Lys 105	Glu	Tyr	Glu	Asp	Met 110	Ile	Ala
•	Thr	Arg	Cys 115	Thr	Cys	Arg										
15	(2) INFO	RMAT:	ION	FOR	SEQ	ID N	0:7:									
	(i)	(B (C) LE) TY) ST	ngth PE: Rand	: 12 amin EDNE	2 am o ac SS:	ino id sing	acid	S							
20		(D) TO	POLO	GY:	line	ar									
	(ii)	MOL	ECUL	E TY	PE:	prot	ein									
	(vii)			TE S ONE:												
25	(ix)) NA	: ME/K CATI												
	(xi)	SEQ	UENC	E DE	SCRI	PTIO	n: s	EQ I	D NO	:7:		:				
	Pro	Arg	Arg	, Asp	Ala 5	Glu	Pro	Val	Leu	Gly	Gly	Gly	Pro	Gly	Gly	Ala
	1				٦.					1 (1					17	

	Cys Arg Ala Arg Arg Leu Tyr Val Ser Phe Arg Glu Val Gly Trp H 20 25 30	is													
	Arg Trp Val Ile Ala Pro Arg Gly Phe Leu Ala Asn Tyr Cys Gln G 35 40 45	ly													
5	Gin Cys Ala Leu Pro Val Ala Leu Ser Gly Ser Gly Gly Pro Pro A 50 55 60	.la													
	Leu Asn His Ala Val Leu Arg Ala Leu Met His Ala Ala Ala Pro 65 70 75 8	1y 10													
10	Ala Ala Asp Leu Pro Cys Cys Val Pro Ala Arg Leu Ser Pro Ile S 85 90 95	er .													
	Val Leu Phe Phe Asp Asn Ser Asp Asn Val Val Leu Arg Gln Tyr (100 105 110	lu													
	Asp Met Val Val Asp Glu Cys Gly Cys Arg 115 120														
15	(2) INFORMATION FOR SEQ ID NO:8:														
20	(2) INFORMATION FOR SEQ ID NO:8: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear														
,	(ii) MOLECULE TYPE: protein														
	(vii) IMMEDIATE SOURCE: (B) CLONE: Vg-1														
25	(ix) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1118														
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:														
	Arg Arg Lys Arg Ser Tyr Ser Lys Leu Pro Phe Thr Ala Ser Asn 1 5 10 15	Ile													

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Cys Lys Lys Arg His Leu Tyr Val Glu Phe Lys Asp Val Gly Trp Gln 30 20 Asn Trp Val Ile Ala Pro Gln Gly Tyr Met Ala Asn Tyr Cys Tyr Gly 45 40 35 Glu Cys Pro Tyr Pro Leu Thr Glu Ile Leu Asn Gly Ser Asn His Ala 5 50 55 Ile Leu Gln Thr Leu Val His Ser Ile Glu Pro Glu Asp Ile Pro Leu 70 75 Pro Cys Cys Val Pro Thr Lys Met Ser Pro Ile Ser Met Leu Phe Tyr 90 10 Asp Asn Asn Asp Asn Val Val Leu Arg His Tyr Glu Asn Met Ala Val 105 Asp Glu Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:9: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Vgr-1 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9: Arg Val Ser Ser Ala Ser Asp Tyr Asn Ser Ser Glu Leu Lys Thr Ala

Cys Arg Lys His Glu Leu Tyr Val Ser Phe Gln Asp Leu Gly Trp Gln 30 20 Asp Trp Ile Ile Ala Pro Lys Gly Tyr Ala Ala Asn Tyr Cys Asp Gly 45 40 35 Glu Cys Ser Phe Pro Leu Asn Ala His Met Asn Ala Thr Asn His Ala - 5 50 Ile Val Gln Thr Leu Val His Leu Met Asn Pro Glu Tyr Val Pro Lys 75 70 Pro Cys Cys Ala Pro Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe 90 10 85 Asp Asp Asn Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn Met Val Val 105 110 100 Arg Ala Cys Gly Cys His 115 (2) INFORMATION FOR SEQ ID NO:10: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: OP-1 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Arg Met Ala Asn Val Ala Glu Asn Ser Ser Ser Asp Gln Arg Gln Ala

10

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	Cys	Lys	Lys	His 20	Glu	Leu	Tyr	Val	Ser	Phe	Arg	Asp	Leu	Gly 30	Trp	Gln
	Asp	Trp	11e 35	Ile	Ala	Pro	Glu	Gly 40	Tyr	Ala	Ala	Tyr	Tyr 45	Cys	Glu	Gly
5	Glu	Cys 50	Ala	Phe	Pro	Leu	Asn 55	Ser	Tyr	Met	Asn	Ala 60	Thr	Asn	His	Ala
	11e 65	Val	Gln	Thr	Leu	Va1 70	His	Phe	Ile	Asn	Pro 75	Glu	Thr	Val	Pro	Lys 80
0	Pro	Cys	Cys	Ala	Pro 85	Thr	Gln	Leu	Asn	Ala 90	Ile	Ser	Val	Leu	Tyr 95	Phe
	Asp	Asp	Ser	Ser 100	Asn	Val	Ile	Leu	Lys 105	Lys	Tyr	Arg	Asn	Met 110		Val
	Arg	Ala	Cys 115	Gly	Cys	His										
15	(2) INFO	RMAT	ION	FOR	SEQ	ID N	0:11	:								
20	(i)	(A (B (C) LE) TY ;) ST	ngth PE: Rand	ARAC : 11 amin EDNE GY:	8 am o ac SS:	ino id sing	acid	S							
	(ii)	-			PE:											
	(vii)				OURC BMF											
25	(ix)		A) NA	ME/K	KEY:											
	(xi)	SEC	QUENC	CE DE	ESCRI	PTIC))))	EQ 1	D NC):11:	:					
	Arį	g Met	Se 1	: Sei	r Val	l Gly	y Ası	ту1	Ası	Thi	Sei	Glu	ı Glı	ı Lys	G1r 15	Ala

5

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	Cys	Lys	Lys	His 20	Glu	Leu	Tyr	Val	Ser 25	Phe	Arg	Asp	Leu	Gly 30	Trp	Gln
	Asp	Trp	Ile 35	Ile	Ala	Pro	Glu	G1y 40	Tyr	Ala	Ala	Phe	Tyr 45	Cys	Asp	Gly
5	Glu	Cys 50	Ser	Phe	Pro	Leu	Asn 55	Ala	His	Met	Asn	Ala 60	Thr	Asn	His	Ala
	Ile 65	Val	Gln	Thr	Leu	Val 70	His	Leu	Met	Phe	Pro 75	Asp	His	Val	Pro	Lys 80
10	Pro	Cys	Cys	Ala	Pro 85	Thr	Lys	Leu	Asn	Ala 90	Ile	Ser	Val	Leu	Tyr 95	Phe
	Asp	Asp	Ser	Ser 100		Val	Ile	Leu	Lys 105		Tyr	Arg	Asn	Met 110	Val	Val
	Arg	Ser	Cys 115	_	Cys	His										
15	(2) INFO	RMAT	ION	FOR	SEQ	ID N	0:12	:								
	(2) INFORMATION FOR SEQ ID NO:12: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear															
20	(ii)	MOI														٠.
	(vii)			ATE S												
25	(ix)	•	A) NA	E: AME/I DCATI												
	(xi) SE	QUEN	CE D	ESCR	IPTI(ON:	SEQ	ID N	0:12	:					
	Se	r Pr	o As	n As	n Va	1 Pr	o Le	u Le	u Gl	u Pr	o Me	t Gl	u Se	r Th	r Ar	g Ser

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	Cys	Gln	Met	Gln 20	Thr	Leu	Tyr	Ile	Asp 25	Phe	Lys	Asp	Leu	Gly 30	Trp	His
	Asp	Trp	Ile 35	Ile	Ala	Pro	Glu	G1y 40	Tyr	Gly	Ala	Phe	Tyr 45	Cys	Ser	Gly
5	Glu	Cys 50	Asn	Phe	Pro	Leu	Asn 55	Ala	His	Met	Asn	Ala 60	Thr	Asn	His	Ala
	Ile 65	· Val	Gln	Thr	Leu	Va1 70	His	Leu	Leu	G1u	Pro 75	Lys	Lys	Val	Pro	Lys 80
10	Pro	Cys	Cys	Ala	Pro 85	Thr	Arg	Leu	Gly	Ala 90	Leu	Pro	Val	Leu	Tyr 95	His
	Lev	. Asn	Asp	Glu 100	Asn	Val	Asn	Leu	Lys 105	Lys	Tyr	Arg	Asn	Met 110	Ile	Val
	Lys	s Ser	Cys 115	-	Cys	His										
15	(2) INFO	RMAT	ION	FOR	SEQ	ID N	0:13	:								
20	(2) INFORMATION FOR SEQ ID NO:13: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 117 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear															
	(ii)) MOI	ECUL	E TY	PE:	prot	ein									
	(vii)) IMM (E	EDIA 3) CL							-						
25	(ix		ATURE A) NA B) LC	ME/K												
	(xi) SEC	QUENC	E DE	SCRI	PTIC)N: S	EQ I	D NC	:13:						
	Gl.	u Lys	s Arg	g Glr	ı Ala	Lys	His	: Lys	Glr	Arg	; Lys	Arg	, Leu	Lys	Ser	Ser

Cys Lys Arg His Pro Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn 25 20 Asp Trp Ile Val Ala Pro Pro Gly Tyr His Ala Phe Tyr Cys His Gly 40 35 Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala 5 55 Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys Ala 70 Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp 90 10 Glu Asn Glu Lys Val Val Leu Lys Asn Tyr Gln Asp Met Val Val Glu 110 105 100 Gly Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:14:

- 15
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 117 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
- 20 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: BMP-4
 - (ix) FEATURE:
- 25 (A) NAME/KEY: Protein
 - (B) LOCATION: 1..117
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys Asn Lys Asn 15 10 1

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	Cys	Arg	Arg	His 20	Ser	Leu	Tyr	Val	Asp 25	Phe	Ser	Asp	Val	Gly 30	Trp	Asn
	Asp	Trp	Ile 35		Ala	Pro	Pro	Gly 40	Tyr	G1n	Ala	Phe	Tyr 45	Cys	His.	Gly
5	Asp	Cys 50	Pro	Phe	Pro	Leu	Ala 55	Asp	His	Leu	Asn	Ser 60	Thr	Asn	His	Ala
	Ile 65	Val	Gln	Thr	Leu	Val 70	Asn	Ser	Val	Asn	Ser 75	Ser	Ile	Pro	Lys	Ala 80
10	Cys	Cys	Val	Pro	Thr 85	Glu	Leu	Ser	Ala	Ile 90	Ser	Met	Leu	Tyr	Leu 95	Asp
	Glu	Tyr	Asp	Lys 100	Val	Val	Leu	Lys	Asn 105	Tyr	Gln	Glu	Met	Val 110	Val	Glu
	Gly	Cys	Gly 115	-	Arg											
15	(2) INFO	RMAT	ION	FOR	SEQ	ID N	0:15	;								
20	(1)	(B	UENC) LE) TY) ST	ngth PE: Rand	: 11 amin EDNE	8 am to ac SS:	ino id sing	acid	is							
	(ii)	MOI	ECUL.	E TY	PE:	prot	ein									
•	(vii)		EDIA 3) CI													
25	(ix)		ATURE A) NA B) LO	ME/F												
	(xi)) SEC	QUENC	CE DI	ESCR:	IPTIC	วพ : . ร	SEQ :	ID NO	0:15	•					
	Ly: 1	s Arį	g Hi:	s Ala	a Ar	g Ar	g Pro	Th:	r Ar	g Ar 10	g Ly:	s Ası	n Hi	s As _l	As ₁	Thr

Cys Arg Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asp 25 20 Asp Trp Ile Val Ala Pro Leu Gly Tyr Asp Ala Tyr Tyr Cys His Gly 40 35 Lys Cys Pro Phe Pro Leu Ala Asp His Phe Asn Ser Thr Asn His Ala 5 55 Val Val Gln Thr Leu Val Asn Asn Met Asn Pro Gly Lys Val Pro Lys 70 Ala Cys Cys Val Pro Thr Gln Leu Asp Ser Val Ala Met Leu Tyr Leu 95 90 . 10 Asn Asp Gln Ser Thr Val Val Leu Lys Asn Tyr Gln Glu Met Thr Val 105 Val Gly Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:16: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 119 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: BMP-3 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..119

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Gln Thr Leu Lys Lys Ala Arg Arg Lys Gln Trp Ile Glu Pro Arg Asn

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Cys Ala Arg Arg Tyr Leu Lys Val Asp Phe Ala Asp Ile Gly Trp Ser 20 25 Glu Trp Ile Ile Ser Pro Lys Ser Phe Asp Ala Tyr Tyr Cys Ser Gly 35 Ala Cys Gln Phe Pro Met Pro Lys Ser Leu Lys Pro Ser Asn His Ala 5 Thr Ile Gln Ser Ile Val Arg Ala Val Gly Val Val Pro Gly Ile Pro 70 75 Glu Pro Cys Cys Val Pro Glu Lys Met Ser Ser Leu Ser Ile Leu Phe 90 10 Phe Asp Glu Asn Lys Asn Val Val Leu Lys Val Tyr Pro Asn Met Thr 105 Val Glu Ser Cys Ala Cys Arg 115 15 (2) INFORMATION FOR SEQ ID NO:17: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 115 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: MIS (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..115 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17: Pro Gly Arg Ala Gln Arg Ser Ala Gly Ala Thr Ala Ala Asp Gly Pro 10

	Cys Ala Leu Arg Glu Leu Ser Val Asp Leu Arg Ala Glu Arg Ser Val 20 25 30														
	Leu Ile Pro Glu Thr Tyr Gln Ala Asn Asn Cys Gln Gly Val Cys Gly 35 40 45														
5	Trp Pro Gln Ser Asp Arg Asn Pro Arg Tyr Gly Asn His Val Val Leu 50 55 60														
	Leu Leu Lys Met Gln Ala Arg Gly Ala Ala Leu Ala Arg Pro Pro Cys 65 70 75 80														
10	Cys Val Pro Thr Ala Tyr Ala Gly Lys Leu Leu Ile Ser Leu Ser Glu 85 90 95														
	Glu Arg Ile Ser Ala His His Val Pro Asn Met Val Ala Thr Glu Cys 100 105 110														
	Gly Cys Arg 115														
15	(2) INFORMATION FOR SEQ ID NO:18:														
20	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 121 amino acids(B) TYPE: amino acid(C) STRANDEDNESS: single														
20	(D) TOPOLOGY: linear														
	(ii) MOLECULE TYPE: protein														
	(vii) IMMEDIATE SOURCE: (B) CLONE: Inhibin alpha														
25	(ix) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1121														
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:														
	Leu Arg Leu Leu Gln Arg Pro Pro Glu Glu Pro Ala Ala His Ala Asn 1 5 10 15														

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Cys His Arg Val Ala Leu Asn Ile Ser Phe Gln Glu Leu Gly Trp Glu 30 20 25 Arg Trp Ile Val Tyr Pro Pro Ser Phe Ile Phe His Tyr Cys His Gly 40 Gly Cys Gly Leu His Ile Pro Pro Asn Leu Ser Leu Pro Val Pro Gly 5 55 Ala Pro Pro Thr Pro Ala Gln Pro Tyr Ser Leu Leu Pro Gly Ala Gln 70 Pro Cys Cys Ala Ala Leu Pro Gly Thr Met Arg Pro Leu His Val Arg 10 Thr Thr Ser Asp Gly Gly Tyr Ser Phe Lys Tyr Glu Thr Val Pro Asn 100 105 Leu Leu Thr Gln His Cys Ala Cys Ile 120 115 (2) INFORMATION FOR SEQ ID NO:19: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 121 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Inhibin betaA (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..121 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19: Arg Arg Arg Arg Gly Leu Glu Cys Asp Gly Lys Val Asn Ile Cys 10 5

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Cys Lys Lys Gln Phe Phe Val Ser Phe Lys Asp Ile Gly Trp Asn Asp 30 20 Trp Ile Ile Ala Pro Ser Gly Tyr His Ala Asn Tyr Cys Glu Gly Glu 40 Cys Pro Ser His Ile Ala Gly Thr Ser Gly Ser Ser Leu Ser Phe His 5 55 Ser Thr Val Ile Asn His Tyr Arg Met Arg Gly His Ser Pro Phe Ala 70 Asn Leu Lys Ser Cys Cys Val Pro Thr Lys Leu Arg Pro Met Ser Met 90 10 -Leu Tyr Tyr Asp Asp Gly Gln Asn Ile Ile Lys Lys Asp Ile Gln Asn Met Ile Val Glu Glu Cys Gly Cys Ser 115 120 (2) INFORMATION FOR SEQ ID NO:20: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Inhibin betaB (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..120 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20: Arg Ile Arg Lys Arg Gly Leu Glu Cys Asp Gly Arg Thr Asn Leu Cys 10 5

	•	Cys	Arg	Gln	G1n 20	Phe	Phe	Ile	Asp	Phe 25	Arg	Leu	Ile	Gly	Trp 30	Asn	Asp
		Trp	Ile	Ile 35	Ala	Pro	Thr	Gly	Tyr 40	Tyr	Gly	Asn	Tyr	Cys 45	Glu	Gly	Ser
5	,	Cys	Pro 50	Ala	Tyr	Leu	Ala	Gly 55	Val	Pro	Gly	Ser	Ala 60	Ser	Ser	Phe	His
		Thr 65	Ala	Val	Val	Asn	Gln 70	Tyr	Arg	Met	Arg	Gly 75	Leu	Asn	Pro	Gly	Thr 80
10		Val	Asn	Ser	Cys	Cys 85	Ile	Pro	Thr	Lys	Leu 90	Ser	Thr	Met	Ser	Met 95	Leu
		Tyr	Phe	Asp	Asp 100	Glu	Tyr	Asn	Ile	Val 105	Lys	Arg	Asp	Val	Pro 110	Asn	Met
		Ile	Val	Glu 115	Glu	Cys	Gly	Cys	Ala 120								
15	(2) I	NFO	RMAT	ION	FOR	SEQ	ID N	0:21	:								
20		(i)	(A (B (C) LE) TY) ST	E CH NGTH PE: RAND POLO	: 11 amin EDNE	4 am o ac SS:	ino id sing	acid	s							
	((ii)	MOL	ECUL	E TY	PE:	prot	ein									
	(1	vii)			TE S ONE:			:a1									
25		(ix)	(A		: ME/K CATI												
		(xi)	SEC	UENO	E DE	ESCRI	PTIC	on: s	EQ 1	D NO	0:21	•					
		Arg	, Arg	g Ala	a Let	ı Asp 5	Th:	. Ası	n Tyi	c Cys	s Phe	e Sei	. Sei	Thi	c Glu	ı Lys 15	s Ası

Cys Cys Val Arg Gln Leu Tyr Ile Asp Phe Arg Lys Asp Leu Gly Trp 20

Lys Trp Ile His Glu Pro Lys Gly Tyr His Ala Asn Phe Cys Leu Gly 45

Pro Cys Pro Tyr Ile Trp Ser Leu Asp Thr Gln Tyr Ser Lys Val Leu 50

Ala Leu Tyr Asn Gln His Asn Pro Gly Ala Ser Ala Ala Pro Cys Cys 65

Val Pro Gln Ala Leu Glu Pro Leu Pro Ile Val Tyr Tyr Val Gly Arg 90

Lys Pro Lys Val Glu Gln Leu Ser Asn Met Ile Val Arg Ser Cys Lys 110

Cys Ser

- 15 (2) INFORMATION FOR SEQ ID NO:22:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 114 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
- 20 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: TGF-beta2
 - (ix) FEATURE:
- 25 (A) NAME/KEY: Protein
 - (B) LOCATION: 1..114
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Lys Arg Ala Leu Asp Ala Ala Tyr Cys Phe Arg Asn Val Gln Asp Asn 1 5 10 15

Cys Cys Leu Arg Pro Leu Tyr Ile Asp Phe Lys Arg Asp Leu Gly Trp 20 25 Lys Trp Ile His Glu Pro Lys Gly Tyr Asn Ala Asn Phe Cys Ala Gly 40 Ala Cys Pro Tyr Leu Trp Ser Ser Asp Thr Gln His Ser Arg Val Leu 5 55 Ser Leu Tyr Asn Thr Ile Asn Pro Glu Ala Ser Ala Ser Pro Cys Cys 75 Val Ser Gln Asp Leu Glu Pro Leu Thr Ile Leu Tyr Tyr Ile Gly Lys 10 Thr Pro Lys Ile Glu Gln Leu Ser Asn Met Ile Val Lys Ser Cys Lys 105 Cys Ser (2) INFORMATION FOR SEQ ID NO:23: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 114 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: TGF-beta3 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..114 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23: Lys Arg Ala Leu Asp Thr Asn Tyr Cys Phe Arg Asn Leu Glu Glu Asn 5 10

WO 94/15966

Cys Cys Val Arg Pro Leu Tyr Ile Asp Phe Arg Gln Asp Leu Gly Trp 30 Cys Ser Gly 45

Lys Trp Val His Glu Pro Lys Gly Tyr Tyr Ala Asn Phe Cys Ser Gly 45

Pro Cys Pro Tyr Leu Arg Ser Ala Asp Thr Thr His Ser Thr Val Leu Sor 70 Cys 75

Cly Leu Tyr Asn Thr Leu Asn Pro Glu Ala Ser Ala Ser Pro Cys Roo Val Pro Gln Asp Leu Glu Pro Leu Thr Ile Leu Tyr Tyr Val Gly Arg 95

Thr Pro Lys Val Glu Gln Leu Ser Asn Met Val Val Lys Ser Cys Lys 100

15 (2) INFORMATION FOR SEQ ID NO:24:

Cys Ser

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
- 20 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (vii) IMMEDIATE SOURCE:
 - (B) CLONE: TGF-beta4
 - (ix) FEATURE:
- 25 (A) NAME/KEY: Protein
 - (B) LOCATION: 1..116
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Arg Arg Asp Leu Asp Thr Asp Tyr Cys Phe Gly Pro Gly Thr Asp Glu
1 5 10 15

	Lys	Asn	Cys	Cys 20	Val	Arg	Pro	Leu	Tyr 25	Ile	Asp	Phe	Arg	Lys 30	Asp	Leu
	Gln	Trp	Lys 35	Trp	Ile	His	Glu	Pro 40	Lys	Gly	Tyr	Met	Ala 45	Asn	Phe	Cys
5	Met	Gly 50	Pro	Cys	Pro	Tyr	Ile 55	Trp	Ser	Ala	Asp	Thr 60	Gln	Tyr	Thr	Lys
	Val 65	Leu	Ala	Leu	Tyr	Asn 70	Gln	His	Asn	Pro	Gly 75	Ala	Ser	Ala	Ala	Pro 80
10	Cys	Cys	Val	Pro	Gln 85	Thr	Leu	Asp	Pro	Leu 90	Pro	Ile	Ile	Tyr	Tyr 95	Val
	Gly	Arg	Asn	Val 100	Arg	Val	Glu	Gln	Leu 105	Ser	Asn	Met	Val	Val 110	Arg	Ala
	Cys	Lys	Cys 115							•						
15	(2) INFO	RMAT	ION	FOR	SEQ	ID N	0:25	:								•
20	(1)	(B) LE 5) TY 5) ST	E CH NGTH PE: RAND	: 11 amin EDNE	4 am o ac SS:	ino id sing	acid	s							
	(ii)	MOL	.ECUL	E TY	PE:	prot	ein									
	(vii)			TE S ONE:			a5									
25	(ix)		A) NA	E: ME/K CATI				·								
	(xi)) SEC	QUENC	CE DE	ESCRI	PTIC	ON: 5	SEQ :	ID NO):25:						
	Lys 1	s Arg	g Gly	y Val	l Gl ₃	y Gli	n Gli	и Туг	r Cys	Phe 10	e Gly	/ Ast	n Ası	ı Gly	y Pro) Asr

5

10

25

-65-

Cys Cys Val Lys Pro Leu Tyr Ile Asn Phe Arg Lys Asp Leu Gly Trp 25

Lys Trp Ile His Glu Pro Lys Gly Tyr Glu Ala Asn Tyr Cys Leu Gly 45

Asn Cys Pro Tyr Ile Trp Ser Met Asp Thr Gln Tyr Ser Lys Val Leu 50

Ser Leu Tyr Asn Gln Asn Asn Pro Gly Ala Ser Ile Ser Pro Cys Gys 80

Val Pro Asp Val Leu Glu Pro Leu Pro Ile Ile Tyr Tyr Val Gly Arg 90

Thr Ala Lys Val Glu Gln Leu Ser Asn Met Val Val Arg Ser Cys Asn

105

Cys Ser

15 (2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:

100

(A) LENGTH: 454 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

20 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vii) IMMEDIATE SOURCE:

(B) CLONE: HUMAN GDF-9

(ix) FEATURE:

(A) NAME/KEY: Protein

(B) LOCATION: 1..454

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

	Met 1	Ala	Arg	Pro	Asn 5	Lys	Phe	Leu	Leu	Trp 10	Phe	Cys	Cys	Phe	Ala 15	Trp
	Leu	Cys	Phe	Pro 20	Ile	Ser	Leu	Gly	Ser 25	Gln	Ala	Ser	Gly	Gly 30	Glu	Ala
5	Gln	Ile	Ala 35	Ala	Ser	Ala	Glu	Leu 40	Glu	Ser	Gly	Ala	Met 45	Pro	Trp	Ser
	Leu	Leu 50	Gln	His	Ile	Asp	G1u 55	Arg	Asp	Arg	Ala	Gly 60	Leu	Leu	Pro	Ala
10	Leu 65	Phe	Lys	Val	Leu	Ser 70	Val	Gly	Arg	Gly	Gly 75	Ser	Pro	Arg	Leu	Gln 80
	Pro	Asp	Ser	Arg	Ala 85	Leu	His	Tyr	Met	Lys 90	Lys	Leu	Tyr	Lys	Thr 95	Tyr
	Ala	Thr	Lys	Glu 100		Ile	Pro	Lys	Ser 105	Asn	Arg	Ser	His	Leu 110		Asn
15	Thr	Val	Arg 115		Phe	Thr	Pro	Cys 120		Arg	His	Lys	Gln 125		Pro	Gly
	Asp	Gln 130		Thr	G1y	Ile	Leu 135		Ser	: Val	Glu	Leu 140		Phe	Asn	Leu
20	Asp 145	_	Ile	. Thr	Thr	Val 150		. Kís	Leu	ı Leu	Lys 155		Val	Leu	. Leu	Tyr 160
	Ast	ılle	. Ast	n Asr	Ser 165		Ser	Phe	e Ser	Ser 170		. Val	. Lys	Cys	Val 175	
	Ast	ı Let	ı Met	11e		: Glu	Pro	Ly:	s Sei 185	r Ser	Ser	Arg	g Thr	Let 190		Arg
25	Ala	a Pro	ту: 19:		r Phe	Thr	Phe	200		r Glī	n Phe	e Glu	205		y Lys	Lys
	His	s Ly:		p Ile	e Glı	n Ile	21:		l Th	r Sei	r Lev	1 Let 220		n Pro	o Lev	ı Val
30	A1:		r As	n Ly	s Ar	g Ser 230		e Hi	s Me	t Se	r Ile 23:		n Phe	e Th	r Cy	s Me

	Lys	Asp	Gln		Glu 245	His	Pro	Ser	Ala	Gln 250	Asn	Gly	Leu	Phe .	Asn 255	Met
	Thr	Leu	Val	Ser 260	Pro	Ser	Leu	Ile	Leu 265	Tyr	Leu	Asn	Asp	Thr 270	Ser	Ala
, 5	Gln	Ala	Tyr 275	His	Ser	Trp	Tyr	Ser 280	Leu	His	Tyr	Lys	Arg 285	Arg	Pro	Ser
	Gln	Gly 290	Pro	Asp	Gln	Glu	Arg 295	Ser	Leu	Ser	Ala	Tyr 300	Pro	Val	Gly	Glu
10	Glu 305		Ala	Glu	Asp	Gly 310	Arg	Ser	Ser	His	His 315	Arg	His	Arg	Arg	Gly 320
	Gln	Glu	Thr	Val	Ser 325	Ser	Glu	Leu	Lys	Lys 330	Pro	Leu	Gly	Pro	Ala 335	Ser
	Phe	Asn	Leu	Ser 340		Tyr	Phe	Arg	G1n 345		Leu	Leu	Pro	Gln 350	Asn	Glu
15	Cys	Glu	1 Leu 355		Asp	Phe	Arg	360		Phe	Ser	Gln	Leu 365		Trp	Asp
	Ası	370		· Val	. Ala	Pro	His 375		Ty:	Ast	Pro	380		Cys	Lys	Gly
20	As ₁ 38!		s Pro	Arg	g Ala	Va]		y His	: Arş	g Tyr	G1 ₃		Pro	Val	His	Thr 400
	Me	t Va	l Glı	n Ası	405		∓ Ty :	r Glu	ı Ly:	s Let 410		Se1	Ser	. Val	419	Arg
	Pr	o Se	r Cy	s Va:		Ala		s Ty:			Le	u Sei	r Val	L Let 430		r Ile
25	G1	u Pr	o As 43		y Se	r Il	e Al	а Ту 44		s Gl	u Ty	r Gl	u Ası 44	p Met 5	110	e Ala
	Th	r Ly 45	_	s Th	r Cy	s Ar	g									

CLAIMS

- 1. Substantially pure growth differentiation factor-9 (GDF-9) and functional fragments thereof.
- 2. An isolated polynucleotide sequence encoding the GDF-9 polypeptide of claim 1.
- 3. The polynucleotide sequence of claim 2, wherein the polynucleotide is isolated from a mammalian cell.
- 4. The polynucleotide of claim 3, wherein the mammalian cell is selected from the group consisting of mouse, rat, and human cell.
- 5. An expression vector including the polynucleotide of claim 2.
- 6. The vector of claim 5, wherein the vector is a plasmid.
- 7. The vector of claim 5, wherein the vector is a virus.
- 8. A host cell stably transformed with the vector of claim 5.
- 9. The host cell of claim 8, wherein the cell is prokaryotic.
- 10. The host cell of claim 8, wherein the cell is eukaryotic.
- 11. Antibodies reactive with the polypeptide of claim 1 or fragments thereof.
- 12. The antibodies of claim 11, wherein the antibodies are polyclonal.

- 13. The antibodies of claim 11, wherein the antibodies are monoclonal.
- 14. A method of detecting a cell proliferative disorder comprising contacting the antibody of claim 11 with a specimen of a subject suspected of having a GDF-9 associated disorder and detecting binding of the antibody.
- 15. The method of claim 14, wherein the cell proliferative disorder is an ovarian tumor.
- 16. The method of claim 14, wherein the detecting is in vivo.
- 17. The method of claim 16, wherein the antibody is detectably labeled.
- 18. The method of claim 17, wherein the detectable label is selected from the group consisting of a radioisotope, a fluorescent compound, a bioluminescent compound and a chemiluminescent compound.
- 19. The method of claim 14, wherein the detection is in vitro.
- 20. The method of claim 19, wherein the antibody is detectably labeled.
- 21. The method of claim 20, wherein the label is selected from the group consisting of a radioisotope, a fluorescent compound, a bioluminescent compound, a chemoluminescent compound and an enzyme.
- 22. A method of treating a cell proliferative disorder associated with expression of GDF-9, comprising contacting the cells with a reagent which suppresses the GDF-9 activity.

- 23. The method of claim 22, wherein the reagent is an anti-GDF-9 antibody.
- 24. The method of claim 22, wherein the reagent is a GDF-9 antisense sequence.
- 25. The method of claim 22, wherein the cell proliferative disorder is an ovarian tumor.
- 26. The method of claim 22, wherein the reagent which suppresses GDF-9 activity is introduced to a cell using a vector.
- 27. The method of claim 26, wherein the vector is a colloidal dispersion system.
- 28. The method of claim 27, wherein the colloidal dispersion system is a liposome.
- 29. The method of claim 28, wherein the liposome is essentially target specific.
- The method of claim 29, wherein the liposome is anatomically targeted.
- 31. The method of claim 29, wherein the liposome is mechanistically targeted.
- 32. The method of claim 31, wherein the mechanistic targeting is passive.
- 33. The method of claim 31, wherein the mechanistic targeting is active.

- 34. The method of claim 33, wherein the liposome is actively targeted by coupling with a moiety selected from the group consisting of a sugar, a glycolipid, and a protein.
- 35. The method of claim 34, wherein the protein moiety is an antibody.
- 36. The method of claim 35, wherein the vector is a virus.
- 37. The method of claim 36, wherein the virus is an RNA virus.
- 38. The method of claim 37, wherein the RNA virus is a retrovirus.
- 39. The method of claim 38, wherein the retrovirus is essentially target specific.

- 1.7 kb

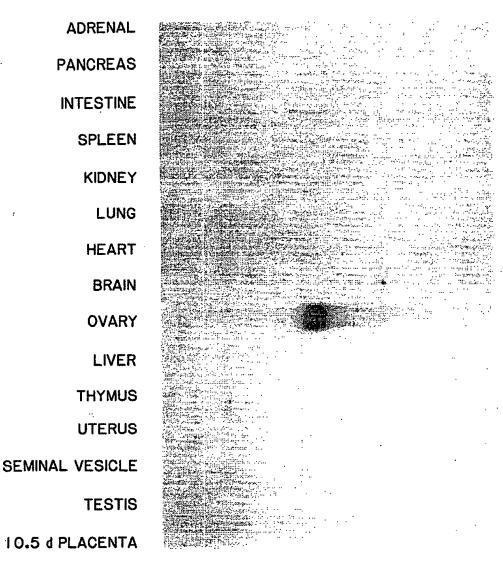


FIG. 1

-	ATGCGTTCCTTCTTCTTCCAAGTCATGGCACTTCCCAGCAACTTCCTGTTGGGGGT	CAACTTCCTGTTGGG	GGT 60	
	MALPS	NFLLGO		
TIGO	TYGCTGCTYTYGCCTGCTGTTTTTCTTAGTAGCCTTTAGCTCTCAGGCTTCTACTGAAGA	TCAGGCTTCTACTGA	AGA 120	
ပ	CCFAWLCFLSSLSS	Q A S. T	ਜ	
ATC.	ATCCCAGAGTGGAGCCAGTGAAAATGTGGAGTCTGAGGCAGACCCCTGGTCCTTGCTGCT	CCCCTGGTCCTTGCT	GCT 180	
ß	QSGASENVESEADPWSLLL	PWSLL	-	
30	GCCTGTAGATGGGACTGACAGGTCTGGCCTCTTGCCCCCCCC	CTTTAAGGTTCTATC	TGA 240	
Д	PVDGTDRSGLLPPLFKVLS	F K V L S	Ω	
IA	TAGGCGAGGTGAGACCCCTAAGCTGCAGCCTCAGAGCACTCTACTACATGAAAA	ACTCTACTACATGAA	AAA 300	
ĸ	RGETPKLQPDSRA	LYYMK	*	2/
S	FCTATAAGACGTATGCTACCAAAGAGGGGGTTCCCAAACC	CAGCAGAAGTCACCT	CTA 360	15
H	LYKTYATKEGVPKPSRSHLY	SRSHL	>)
A.	CAATACCGTCCGGCTCTTCAGTCCCTGTGCCCAGCAGAGCAGGCACCCAGCAACCAGGT	GGCACCCAGCAACCA	GGT 420	
Z	NTVRLFSPCAQQEQAP	APSNQV	>	
3A	GACAGGACCGCTGCCGATGGTGGACCTGCTGTTTAACCTGGACCGGGTGACTGCCATGGA	CCGGGTGACTGCCAT	GGA 480	
H	GPLPMVDLLFNLD	R V T A M	ы	
S	ACACTTGCTCAAATCGGTCTTGCTATACACTCTGAACAACTCTGCCTCTTCCTCCTCCTCCAC	тесстсттсстсстс	CAC 540	
Ħ	HLLKSVLLYTLNNS	ASSSS	E	
2	TGTGACCTGTATGTGTGACCTTGTGGTAAAGGAGGCCATGTCTTCTGGCAGGGCACCCCC	TTCTGCCAGGCCACC	CCC 600	
>	VTCMCDLVVKEAMS	S. G. R. A. P.	<u>م</u>	

FIG.2

FIG.2

11	1141	GGACTGTCCTAGGGCGGTCAGGCATCGGTATGGCTCTCCTGTGCACACCATGGTCCAGAA	1200
1	 	DCPRAVRHRYGSPVHTMVQN	
12	1201	TATAATCTATGAGAAGCTGGACCCTTCAGTGCCAAGGCCTTCGTGTGTGCCGGCCAAGTA	1260
		IIYEKLDPSVPRPSCVPGKY	
12	1261	CAGCCCCCTGAGTGTGTTGACCCATTGAACCCGACGGCTCCATCGCTTACAAAGAGTACGA	1320
		SPLSVLTIEPDGSIAYKEYE	
13 U 3	121	ř	1380
BS		DMIATRCTCR *	
1381	81	CTGGCAGAGCAATGCTGTGGGCCTTAGAGTGCCTGGGCAGAGAGCTTCCTGTGACCAGTC	1440
ח 14 14	141	TCTCCGTGCTGCTCAGTGCACACTGTGTGTGGGGGGAAGTGTGTGT	1500
E 15	1501	CATCGAGTGCAGTGTCCGTAGGTGTAAAGGGCCACACTCACT	1560
	1561	GTGAAATGTAACTCATTTGGAGAGCTCTTTCTCCCCACGAGTGTAGTTTTCAGTGGACAG	1620
EE 16	1621	ATTIGITAGCATAAGTCTCGAGTAGAATGTAGCTGTGAACATGTCAGAGTGCTGTGGTTT	1680
•	1681	TATCTGAAGAATAAACTGTTGATGGCAT 1712	

FIG.20

KGFCRNFCHRHOLFINF-QDLGWHKWVIAPKGFMANYCHGECPFSMTTYLNS FLFPQNECELHDFRLSF-SQLKWDNWIVAPHRYNPRYCKGDCPRAVRHRYGS	PREDAEPVIGGGPGGAGRARRLYVSF-REVGWHRWVIAPRGFLANY CYGE GALPVALSGSGGP RRKRSYSKLPFTASNICKRRLYVSF-RDVGWQNWVIAPQGYMANYCYGE GYPLTEILNG- RWASNASDYNSSELKTACRKHELYVSF-QDLGWQDWIIAPRGYAANYCDGE SFPLNAHMNA RWANNAENSSSDORQACKHELYVSF-QDLGWQDWIIAPEGYAANYCDGE SFPLNAHMNA RMSSVGDYNTSEQRQACKHELYVSF-RDLGWQDWIIAPEGYAAFYCDGE SFPLNAHMNA SPNNVPLLEPMESTRSCACKHELYVSF-RDLGWQDWIIAPEGYAAFYCDGE SFPLNAHMNA SPNNVPLLEPMESTRSCACKHELYVSF-RDLGWQDWIIAPEGYAFYCDGE SFPLNAHMNA SPNNVPLLEPMESTRSCACKHPLYVDF-SDVGWNDWIVAPPGYAAFYCGGE SFPLNAHMNA SPNNVPLLEPMESTRSCACKHPLYVDF-SDVGWNDWIVAPPGYAFYCGGE SFPLADHLINS EKRQAKHKQRKRLKSSCKRHPLYVDF-SDVGWNDWIVAPPGYAFYCGGE SFPLADHLINS RSPKHHSQRARKKNKNCRRHSLYVDF-SDVGWNDWIVAPPGYAFYCGGE SFPLADHLINS RRAPARRYQMIEPRN CARRYLKVDF-ADIGWSEWIISPKSFDAYY SGACFPPLADHLINS PGRAQRSAGATAADGPALRELSVDLRAERSVLIPPKGYDAYY CHGC GLHIPPNLSLPV RRRARRGLECDGRYNICKRQFFVSF-KDIGWNEWIIYPPGSYHANYCEGE SPAYLAGVPGSAS RRALDTNYCFRNVDNCCKRQFYSIDFRDLGWK-WIHEPKGYNANF CAGGEPYIWSSD RRALDTNYCFRNVEENCYRPLYIDFRRDLGWK-WIHEPKGYNANF CAGGEPYIWSAD RRALDTNYCFRNVEENCYRPLYIDFRRDLGWK-WIHEPKGYNANF CAGGEPYIWSAD RRDLDTDYCFGBGTDEKN CVRPLYIDFRRDLGWK-WIHEPKGYNANF CAGGEPYIWSAD RRDLDTDYCFGRONGPNCVRPLYIDFRRDLGWK-WIHEPKGYNANF CAGGEPYIWSMD	
KRRAAISVPKGFCRNFCHI FNLSEYFKQFLFPQNECEI	PREDAEPVLGGGPGGACRU RRKRSYSKLPFTASNICKI RVSSASDYNSSELKTACKI RVSSASDYNSSELKTACKI RWANVAENSSDQRQACKI SPNNVPLLEPMESTRSQKI EKRQAKHKQRKRLKSSCKI RSPKHHSQRARKKNKNCRI KRHARRPTRRKNHDDTCRI QTLKKARRKQWIEPRNCAI PGRAQRSAGATAADGPCAI LRLLQRPPEEPAAHANCHI RRRRRGLECDGKVNICCI RICKGLECDGKYNICCI RRALDTNYCFSSTEKNCCI KRALDAAYCFRNVQDNCCI KRALDTNYCFSSTEENCCI KRALDTNYCFGPGTDEKNCCI KRALDTNYCFGNNGPNCCI	
GDF-3 GDF-9	GDF-1 Vg-1 Vg-1 OP-1 BMP-5 60A BMP-2 BMP-4 DPP BMP-3 MIS Inhibin &A Inhibin &B TGF-&1 TGF-&3 TGF-&3 TGF-&3	

FIG.3a

500000	<u> </u>	∝	H	II.	T (6):	II G	H (6):	Y.	K O	K O	K	X	H	S	<u></u>	S	S	S	S	S
arrest.	ΟH	ď	Ŏ	Ö	Ö	Ŏ	Ö	Ŭ	Ö	Ŏ	đ	Ö	4	Ů.	Ö	Ÿ.	Y	Ϋ́	Ÿ.	2_
88888	SDKNVILRHYEDMVVDE	NSDNVVLRQYEDMVVDEC	TLVHSIEPEDIPLPCOVPTKMSPISMLFY-DNNDNVVLRHYENMAVDEC	TLVHLMNPEYVPKPCAPTKLNAISVLYF-DDNSNVILKKYRNMVVRA	PTQLNAISVLYF-DDSSNVILKKYRNMVVRAC	PTKLNAISVLYF-DDSSNVILKKYRNMVVRS	PTRLGAL PVLYH - LNDENVNLKKYRNMIVKS	PTELSAISMLYL-DENEKVVLKNYQDMVVEG	PTELSAISMLYL-DEYDKVVLKNYQEMVVEC	PTQLDSVAMLYL-NDQSTVVLKNYQEMTVVGCC	PEKMSSLSILFF-DENKNVVLKVYPNMTVESGP	ERISAHHVPNMVATE	GGYSFKYETVPNLLTQH	NHYRMRGHSPFANLKS CVPTKLRPMSMLYY-DDGQNIIKKDIQNMIVEE CCS	DEYNIVKRDVPNMIVEE	GRKPKV-EQLSNMIVRS	SQDLEPLTILYY-IGKTPKI-EQLSNMIVKSTK	GRTPKV-EQLSNMVVKS	PQTLDPLPIIYY-VGRNVRV-EQLSNMVVRAGIGS	PDVLEPLPIIYY-VGRTAKV-EQLSNMVVRSCNGS
0000	VPTKLSPISMLYQ-D: VPGKYSPLSVLTI-E)	VPARLSPISVLFF-D	VPTKMSPISMLFY-DI	APTKLNAISVLYF-D	APTQLNAISVLYF-D	APTKLNAISVLYF-D	APTRLGALPVLYH-L	VPTELSAISMLYL-D	1	!	VPEKMSSLSILFF-D	VPTAYAGKLLISLSE	AALPGTMRPLHVRTTSD	VPTKLRPMSMLYY-DI	I PTKLSTMSMLYF-DI	VPOALEPLPIVYY-V	ŀ	VPODLEPLTILYY-V	VPQTLDPLPIIYY-V	-
	> 7	ď	ŭ	ŭ	ŭ	ŭ	ŭ	$\ddot{\mathbf{o}}$	ŏ	ŏ	ŭ	$\check{\mathbf{o}}$	ŏ	ŏ	ŏ	ŏ	Ŏ,	ŭ	Ŏ	Ŭ
	SNYAFWQALMHMADPKVPKAVCVPTKLSPISMLYQ-DSDKNVILRHYEDMVVDE CC PVHTMVQNIIYEKLDPSVPRPSCVPGKYSPLSVLTI-EPDGSIAYKEYEDMIATRCTIR	PALINHAVLRALIMHA AAPGAADLPCCV PARLSPISVLFF - DNSDNVVLRQYEDMVVDE	SNHAILOTLVHSIEPEDIPLE	TNHAIVQTLVHLMNPEXVPKP	TNHAIVQTLVHFINPETVPKPCA-	TINHAIVOTLVHLMFPDHVPKPCA	TNHAIVQTLVHLLEPKKVPKPCA	TNHAIVQTLVNSVNSKIPKACOV	TNHAIVQTLVNSVNSSIPKACOV	TNHAVVQTLVNNMNPGKVPKACOV	SNHATIQSIVRA-VGVVPGIPEPCOV	GNHVVLLLKMQARGAALARPPCOVPTAYAGKLLISLSEERISAHHVPNMVATE	pgapptpaopysllpgaopcalpgtmrplhyrttsdggysfkyetvpnlltohga	SFHSTVI	SFHTAW	TQYSKVLALYNQHNPGASAAF	TOHSRVLSLYNTINPEASASPCOV	THSTVLGLYNTLNPEASASPCOVPODLEPLTILYY-VGRTPKV-EQLSNMVVKS	TOYTKVLALYNQHNPGASAAFCCV	TOYSKVLSLYNQNNPGASISPECV-
													ಶ	8A	ßB					
	GDF-3 GDF-9	GDF-1	Vg-1	Vgr-1	0P-1	BMP-5	60A	BMP-2	BMP-4	DPP	BMP-3	MIS	Inhibin	Inhibin	Inhibin	TGF-B1	TGF-82	TGF-83	TGF-84	TGF-B5

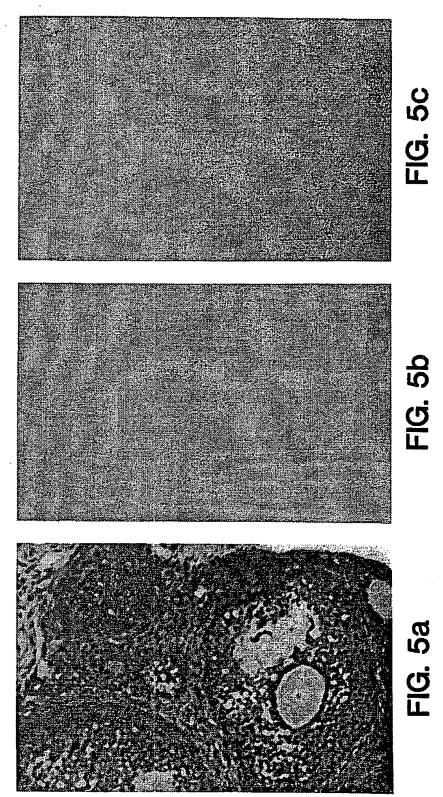
7,	1	
//	1	~

								•			
TGF− <i>β</i> 5	36	25	32	34	37	36	36	36	35	33	35
TGF−β4	33	22	34	32	39	37	36	38	33	32	33
TGF−β3	32	25	33	37	39	38	36	40	36	35	32
TGF- <i>8</i> 2	31	25	32	36	37	38	35	39	34	33	35
TGF− <i>β</i> I	36	23	33	34	35	34	34	38	SE.	34	35
INHIBIN B B	41	31	35	37	41	42	37	39	42	42	42
INHIBIN <i>B</i> A	42	30	37	44	44	43	43	36	42	41	39
іннівін а	25	27	23	22	25	24	24	24	22	22	19
міѕ	22	21	34	30	24	27	24	25	27	27	25
BMP-3	62	73	42	49	44	42	43	41	48	47	43
DPP	47	32	41	48	29	28	57	54	74		100
BMP-4	20	34	43	26	09	58	59	54	92	00	
BMP-2	53	33	42	28	61	09	61	57	00		1
60A	47	30	41	51	7	· 69	7	100	g er to:	itality: I	<u> </u>
BMP-5	20	31	46	99	16	88	100	ı	,	ı	t
OP-1	20	30	47	57	87	100	-	1	,	ı	ı
Vgr-1	53	31	46	58	00		ı	•		ı	ı
Vg-I	57	30	57	100		ı	1 	1	J I	ı	ı
GDF-I	20	27	100	1	ŀ	ı	i	ı	1		ı
GDF-9	33	100	•	ı	1	1	ı	t	ı	ı	ı
GDF-3	100	•	ı	ı	•	ı	1		1	1	ı
	GDF-3	GDF-9	GDF-1	Vg-1	Vgr-1	1-d0	BMP-5	60A	BMP-2	BMP-4	ОРР

FIG.4a

٥	1	1	\Box
a	/	ı	2

TGF- β 5	30	26	24	36	58	82	20	73	79	100	
TGF-β4	27	53	24	33	30	98	89	74	100	1	
TGF-β3	32	25	24	36	37	7.8	82	100	3	1	
TGF- β 2	32	23	22	37	34	74	100	ı	ı	•	
TGF− <i>β</i> Ι	32	28	23	41	35	100	ı	1	1	•	
INHIBIN #B	37	25	25	63	100	1	ı	1	I	(A): (A)(A)	1
INHIBIN &A	36	24	56	100	•	1	1	ı	•		
INHIBIN a	53	18	100	**************************************	1 I	1	•	•	ı	ı	
_ MIS	30	100	1	ı	•	ı	٠	ı	•	ı	
BMP-3	100	-	•	1	ı	ı	1	ı	ı	ŧ	
DPP	1	•	ı	1	1	1	ı	ı	ı	ı	
BMP-4	1	1	1	•	ı	1	1	ı	1	ı	4
BMP-2	. 1	ŧ	1	ı	1	ı	1	1	ı	1	IG.4 k
60A	•	ι	ı	4	ı	f	F	•	1	1	Ī.
BMP-5	1	1	1	ŧ	•	ı	1	1	•	ı	
OP-I	1	i	ı	1	•	1	1	ı	ı	ı	
Vgr- I	ı	ı	ı	•	1	. 1	ı	1	•	i	
Vg- i	ı	ŧ	ŧ	1	1	1	ı	ı	ı	ı	
GDF-I	t	ı	1	1	•	ı	ı	ı	ı	t	
GDF-9	1	ı	ı	•	1	ı	ı	ı	ı	1	
GDF-3	t	ı	1	ı	ı	1	ı	1	ı	1	
	BMP-3	MIS	INHIBIN &	INHIBIN BA	INHIBIN AB	TGF-BI	TGF-82	TGF-#3	TGF-84	TGF-#5	



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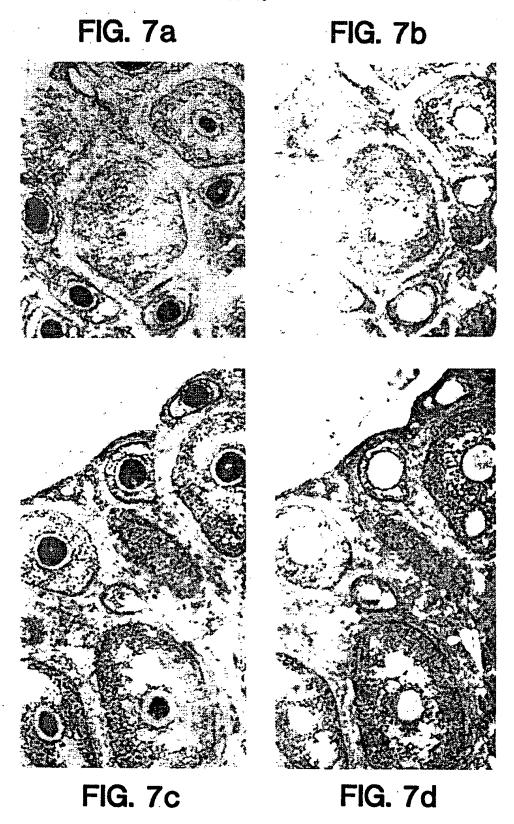
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1		50
51	LPVDGTDRSGLLPPLFKVLSDRRGETPKLQPDSRALYYMKKLYKTYATKE	100
51	QHIDERDRAGLLPALFKVLSVGRGGSPRLQPDSRALHYMKKLYKTYATKE	100
101	GVPKPSRSHLYNTVRLFSPCAQQEQAPSNQVTGPLPMVDLLFNLDRVTAM	150
101	GIPKSNRSHLYNTVRLFTPCTRHKQAPGDQVTGILPSVELLFNLDRITTV	150
151	EHLLKSVLLYTLNNSASSSSTVTCMCDLVVKEAMSSGRAPPRAPYSFTL.	199
151	EHLLKSVLLYNINNSVSFSSAVKCVCNLMIKEPKSSSRTLGRAPYSFTFN	200
200	KKHRWIEIDVTSLLQPLVTSSERSIHLSVNFTCTKDQVPE	239
201	SQFEFGKKHKWIQIDVTSLLQPLVASNKRSIHMSINFTCMKDQLEHPSAQ	250
240	DGVFSMPLSVPPSLILYLNDTSTQAYHSWQSLQSTWRPLQHPGQA.GVAA	288
251	NGLFNMTL.VSPSLILYLNDTSAQAYHSWYSLHYKRRPSQGPDQERSLSA	299
289	RPVKEEATEVERSP. RRRRGQKAIRSEAKGPLLTASFNLSEYFKQFLFP	336
	YPVGEEAAEDGRSSHHRHRRGQETVSSELKKPLGPASFNLSEYFRQFLLP	
	QNECELHDFRLSFSQLKWDNWIVAPHRYNPRYSKGDSPRAVRHRYGSPVH	
	QNEGELHDFRLSFSQLKWDNWIVAPHRYNPRY	
	TMVQNIIYEKLDPSVPRPS VPGKYSPLSVLTIEPDGSIAYKEYEDMIAT	
	TMVQNIIYEKLDSSVPRPS VPAKYSPLSVLTIEPDGSIAYKEYEDMIAT	449
	RETER 441	
450	кетек 454	

^{64%} FIG. 6

90%

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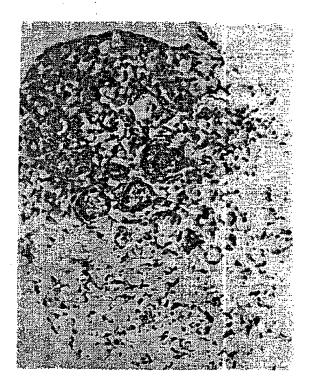


FIG. 8a

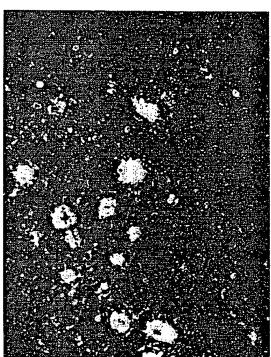
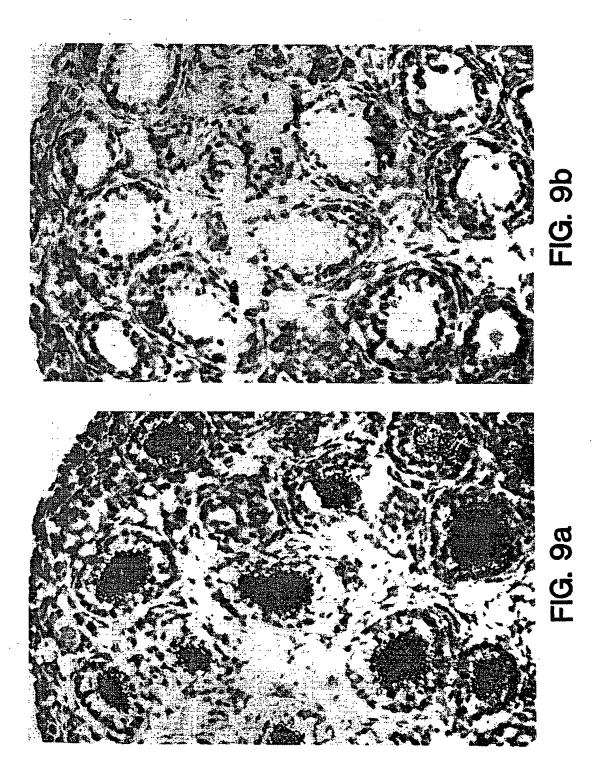
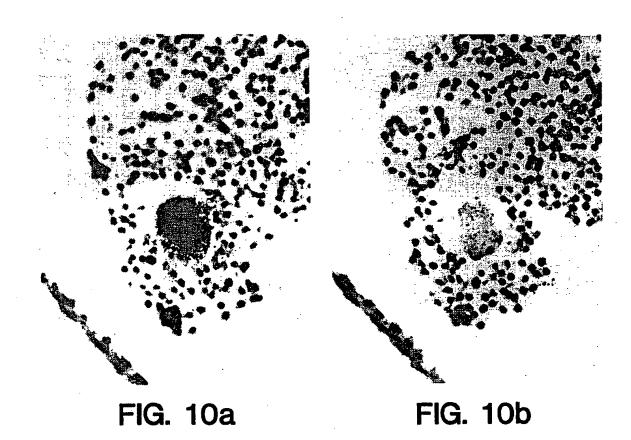


FIG. 8b



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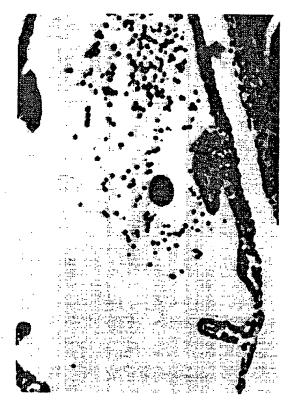


FIG. 11a

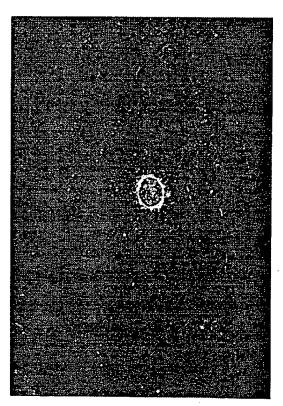


FIG. 11b

INTERNATIONAL SEARCH REPORT

In ational application No. PCT/US94/00685

A. CLA	SSIFICATION OF SUBJECT MATTER								
、,	US CL.: Please See Extra Sheet.								
	According to International Patent Classification (IPC) or to both national classification and IPC								
B. FIELDS SEARCHED									
Minimum documentation searched (classification system followed by classification symbols)									
U.S. : 536/23.5, 23.4; 435/320.1, 69.1, 69.4, 91.1, 91.4, 252.3, 252.33; 530/350, 399, 388.23									
Documentat	Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched								
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)									
	Dialog, APS search terms: growth differentiation factor-9 Sequence search: GenBank, GeneSeq, PIR, SwissPro								
C. DOC	UMENTS CONSIDERED TO BE RELEVANT								
Category*	Citation of document, with indication, where ap	propriate, of the relevant passages	Relevant to claim No.						
A	MOLECULAR ENDOCRINOLOGY, Volume 4, issued 1990, Lee, "Identification of a Novel Member (GDF-1) of the Transforming Growth Factor-beta Superfamily," pages 1034-1039.								
A	PROCEEDINGS OF THE NAT SCIENCES, USA, Volume 88, "Expression of growth/differentiation of a bicis 4250-4254, see entire document.	on factor I in the nervous	1-39						
X Furth	ner documents are listed in the continuation of Box C	. See patent family annex.							
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	cument published prior to the international filing date but later than a priority date claimed	*&* document member of the same patern	t family						
	actual completion of the international search	Date of mailing of the international se	•						
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INTERNATIONAL SEARCH REPORT

h. .ational application No.
PCT/US94/00685

- 1	tion). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
X,P	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 268, Number 5, issued 15 February 1993, McPherron et al., "GDF-3 and GDF-9:Two New Members of the Transforming Growth Factor-beta Superfamily Containing a Novel Pattern of Cysteines", pages 3444-3449, see figure 2.	1-39
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INTERNATIONAL SEARCH REPORT

li sational application No. PCT/US94/00685

US CL: 536/23.5, 23.4; 435/320	0.1, 69.1, 69.4, 91.1, 91.4 , 252.3, 252.33; 530/350, 399	9, 388.23
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